



# Genome Sequence of a Small RNA Virus of the Southern Corn Rootworm, *Diabrotica undecimpunctata howardi* Barber (Coleoptera: Chrysomelidae)

Sijun Liu,<sup>a</sup> Arnubio Valencia-Jiménez,<sup>b,c</sup> Molly Darlington,<sup>c</sup> Ana M. Vélez,<sup>c</sup>  Bryony C. Bonning<sup>d</sup>

<sup>a</sup>Department of Entomology, Iowa State University, Ames, Iowa, USA

<sup>b</sup>Departamento de Producción Agropecuaria, Universidad de Caldas, Manizales, Colombia

<sup>c</sup>Department of Entomology, University of Nebraska—Lincoln, Lincoln, Nebraska, USA

<sup>d</sup>Department of Entomology and Nematology, University of Florida, Gainesville, Florida, USA

**ABSTRACT** The genome sequence of a novel small RNA virus, tentatively named *Diabrotica undecimpunctata virus 1* (DuV1), was discovered from the transcriptome of the southern corn rootworm, *Diabrotica undecimpunctata howardi* Barber. DuV1 has a positive-sense, single-stranded RNA genome that encodes a single polyprotein of 3,401 amino acids with limited similarity to other viruses.

The southern corn rootworm (SCR), *Diabrotica undecimpunctata howardi* Barber, is a polyphagous pest that impacts multiple crops in the southeastern United States, southern Canada, and northern Mexico (1). While several small RNA viruses have been reported from other rootworm species (2–4), this is the first report of a virus genome sequence from the transcriptome of *D. undecimpunctata*.

SCR were supplied by Crop Characteristics (Farmington, MN). Total RNA was isolated from three biological replicates of multiple developmental stages (eggs; first-, second-, and third-instar larvae; pupae; adult females; and adult males) using the RNeasy minikit (Qiagen, Valencia, CA, USA). RNA extractions from each stage-specific sample were used for cDNA library construction with poly(A) selection. The NEBNext Ultra RNA library prep kit for Illumina was used for library preparation. cDNA libraries were paired-end sequenced (150-nucleotide [nt] reads) on a HiSeq 2500 platform (Illumina, San Diego, CA). High-quality sequencing reads (48.2 million reads for males, 41.5 million reads for females; mean quality score, 37.5) were assembled by using the Trinity assembler v2.6.6 (5). Default parameters were used for all software unless otherwise noted. Putative viral sequences were initially identified by BLASTx analysis (6) against a local viral sequence database with the following cutoff values: contig length, >300 nt; protein sequence length, >50 amino acids [aa]; and E value, 0.01. Here, we describe one of two near-complete viral genome sequences identified from this analysis.

A 10,483-nt contig with a G+C content of 47% was assembled from the sequencing reads from both female and male adult SCR and was named *Diabrotica undecimpunctata virus 1* (DuV1). A total of 1,070 reads from males and 1,078 from females mapped to viral sequences with a coverage of 15.3 and 15.4, respectively. Only a few short fragments were identified from larval samples. A 10,203-nt coding region was flanked by a 5' untranslated region (UTR) of at least 139 nt and a 3' UTR of at least 141 nt. The predicted molecular mass of the polyprotein is 378.7 kDa. The N-terminal half of the hypothetical protein contains putative nonstructural proteins, with the C-terminal half comprised of putative capsid proteins. The nonstructural protein sequence contained six conserved domains, four of which overlapped, i.e., Macro\_Poa1p\_like domain (aa 46 to 151; NCBI accession no. [cd02901](https://.ncbi.nlm.nih.gov/nuccore/cd02901)); A1pp, an appr-1"-p processing enzyme (aa 48 to 149; SMART accession no. [SM00506](https://smart.embl.de/smart/show_results.cgi?tbl_function=1&tbl_scan=1&tbl_seq=SM00506)); Macro, an ADP-ribose binding module (aa 89 to

**Citation** Liu S, Valencia-Jiménez A, Darlington M, Vélez AM, Bonning BC. 2020. Genome sequence of a small RNA virus of the southern corn rootworm, *Diabrotica undecimpunctata howardi* Barber (Coleoptera: Chrysomelidae). *Microbiol Resour Announc* 9:e00379-20. <https://doi.org/10.1128/MRA.00379-20>.

**Editor** Kenneth M. Stedman, Portland State University

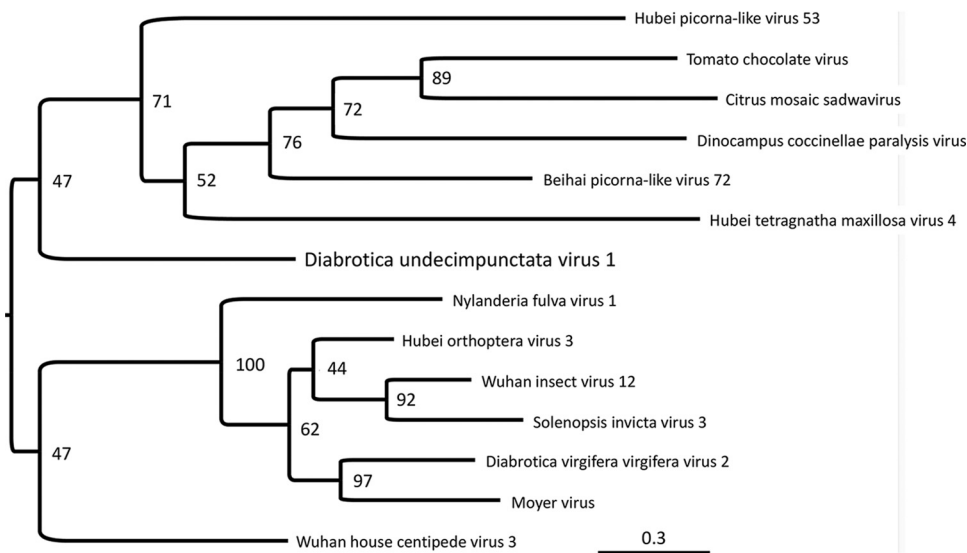
**Copyright** © 2020 Liu et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Bryony C. Bonning, [bbonning@ufl.edu](mailto:bbonning@ufl.edu).

**Received** 10 April 2020

**Accepted** 28 May 2020

**Published** 25 June 2020



**FIG 1** Phylogenetic analysis of DuV1 RdRP. The RdRP sequences identified by an RdRP domain search on BLAST annotation were extracted from the NCBI database. Sequences were aligned by use of MAFFT v7.4.07 with the AUTO strategy. The phylogenetic tree was constructed using the maximum likelihood (ML) method. ModelFinder implemented in IQ-TREE was used to choose the best partitioning scheme and models. ML analysis was performed using IQ-TREE with 10,000 Ultrafast bootstraps (9). Constructed trees were uploaded to Interactive Tree of Life (<http://itol.embl.de>) for visualization and annotation. The selected RNA viruses and their NCBI genome accession numbers are as follows: *Beihai picorna-like virus 72*, [APG76793.1](https://.ncbi.nlm.nih.gov/nuccore/APG76793.1); *Citrus mosaic sadwavirus*, [AWY04239.1](https://.ncbi.nlm.nih.gov/nuccore/AWY04239.1); *Dinocampus coccinellae paralysis virus*, [YP\\_009111311.1](https://.ncbi.nlm.nih.gov/nuccore/YP_009111311.1); *Diabrotica virgifera virgifera virus 2*, [YP\\_009352233.1](https://.ncbi.nlm.nih.gov/nuccore/YP_009352233.1); *Hubei orthoptera virus 3*, [YP\\_009336506.1](https://.ncbi.nlm.nih.gov/nuccore/YP_009336506.1); *Hubei picorna-like virus 53*, [YP\\_009336820.1](https://.ncbi.nlm.nih.gov/nuccore/YP_009336820.1); *Hubei tetragnatha maxillosa virus 4*, [YP\\_009336535.1](https://.ncbi.nlm.nih.gov/nuccore/YP_009336535.1); *Moyer virus*, [AOC55061.1](https://.ncbi.nlm.nih.gov/nuccore/AOC55061.1); *Nylanderia fulva virus 1*, [YP\\_009268643.1](https://.ncbi.nlm.nih.gov/nuccore/YP_009268643.1); *Solenopsis invicta virus 3*, [ACZ65579.1](https://.ncbi.nlm.nih.gov/nuccore/ACZ65579.1); *Tomato chocolate virus*, [ACU01024.1](https://.ncbi.nlm.nih.gov/nuccore/ACU01024.1); *Wuhan house centipede virus 3*, [YP\\_009342325.1](https://.ncbi.nlm.nih.gov/nuccore/YP_009342325.1); and *Wuhan insect virus 12*, [YP\\_009345893.1](https://.ncbi.nlm.nih.gov/nuccore/YP_009345893.1).

149; NCBI accession no. [pfam01661](https://pubmed.ncbi.nlm.nih.gov/1661/)); and Ymd, an O-acetyl-ADP-ribose deacetylase (regulator of RNase III) (aa 63 to 174; NCBI accession no. [COG2110](https://pubmed.ncbi.nlm.nih.gov/2110/)). An RNA\_helicase domain was located between aa 812 and 913 (NCBI accession no. [pfam00910](https://pubmed.ncbi.nlm.nih.gov/910/)), and an RdRP domain was located between aa 1934 and 2367 (EMBL-EBI accession no. [pfam00680](https://pubmed.ncbi.nlm.nih.gov/680/), NCBI accession no. [cd01699](https://pubmed.ncbi.nlm.nih.gov/1699/)). Two cysteine protease sequence motifs, GxCG and GxHxxG, were identified between aa 1734 and 1737 (GDCG) and aa 1755 and 1760 (GIHVAG). No conserved domains were found in the putative viral capsid region.

The genome organization of DuV1 encoding a single polyprotein with N-terminal nonstructural proteins is similar to that of *Diabrotica virgifera virgifera virus 2* (DvV2) (3). However, the highest BLASTp score (6, 7) was an unclassified virus, *Wuhan house centipede virus 3* (GenBank accession no. [YP\\_009342325.1](https://pubmed.ncbi.nlm.nih.gov/1)) (8), with only 36% sequence coverage and 31% amino acid identity. Similar sequences were limited to nonstructural proteins. A phylogenetic tree based on the RdRP sequences of DuV1 and selected related RNA viruses highlights the novelty of this virus (Fig. 1). Based on this analysis, DuV1 represents a novel virus.

**Data availability.** The genome sequence of DuV1 is available in GenBank under accession no. [MN646770](https://pubmed.ncbi.nlm.nih.gov/1). Raw reads for SRA accession no. [SRX8018653](https://pubmed.ncbi.nlm.nih.gov/1) to [SRX8018656](https://pubmed.ncbi.nlm.nih.gov/1) are available at BioProject no. [PRJNA615920](https://pubmed.ncbi.nlm.nih.gov/1).

## ACKNOWLEDGMENTS

This research was supported by the USDA National Institute of Food and Agriculture, Hatch project IOW05450; by State of Iowa funds; University of Nebraska—Lincoln funds; the 2016 DuPont Young Professor Grant awarded to A.M.V.; and by the University of Florida Institute of Food and Agricultural Sciences.

## REFERENCES

- Krysan JL. 1986. Introduction: biology, distribution, and identification of pest *Diabrotica*. In Krysan JLM, Thomas A (ed), *Methods for the study of pest Diabrotica*. Springer, New York, NY.
- Liu S, Chen Y, Sappington TW, Bonning BC. 2017. Genome sequence of a novel positive-sense, single-stranded RNA virus isolated from western corn rootworm, *Diabrotica virgifera virgifera* LeConte. *Genome Announc* 5:e00366-17. <https://doi.org/10.1128/genomeA.00366-17>.
- Liu S, Chen Y, Sappington TW, Bonning BC. 2017. Genome sequence of

- Diabrotica virgifera virgifera* virus 2, a novel small RNA virus of the western corn rootworm, *Diabrotica virgifera virgifera* LeConte. *Genome Announc* 5:e00365-17. <https://doi.org/10.1128/genomeA.00365-17>.
4. Liu S, Chen Y, Sappington TW, Bonning BC. 2017. Genome sequence of the first coleopteran iflavirus isolated from western corn rootworm, *Diabrotica virgifera virgifera* LeConte. *Genome Announc* 5:e01530-16. <https://doi.org/10.1128/genomeA.01530-16>.
  5. Haas BJ, Papanicolaou A, Yassour M, Grabherr M, Blood PD, Bowden J, Couger MB, Eccles D, Li B, Lieber M, MacManes MD, Ott M, Orvis J, Pochet N, Strozzi F, Weeks N, Westerman R, William T, Dewey CN, Henschel R, Leduc RD, Friedman N, Regev A. 2013. De novo transcript sequence reconstruction from RNA-seq using the Trinity platform for reference generation and analysis. *Nat Protoc* 8:1494–1512. <https://doi.org/10.1038/nprot.2013.084>.
  6. Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res* 25:3389–3402. <https://doi.org/10.1093/nar/25.17.3389>.
  7. Altschul SF, Wootton JC, Gertz EM, Agarwala R, Morgulis A, Schaffer AA, Yu YK. 2005. Protein database searches using compositionally adjusted substitution matrices. *FEBS J* 272:5101–5109. <https://doi.org/10.1111/j.1742-4658.2005.04945.x>.
  8. Shi M, Lin XD, Tian JH, Chen LJ, Chen X, Li CX, Qin XC, Li J, Cao JP, Eden JS, Buchmann J, Wang W, Xu J, Holmes EC, Zhang YZ. 2016. Redefining the invertebrate RNA virosphere. *Nature* 540:539–543. <https://doi.org/10.1038/nature20167>.
  9. Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS. 2018. UFBoot2: improving the ultrafast bootstrap approximation. *Mol Biol Evol* 35:518–522. <https://doi.org/10.1093/molbev/msx281>.