



Draft Genome Sequence of *Proteus mirabilis* Strain UMB0038, Isolated from the Female Bladder

Noreen Gallian,^a Taylor Miller-Ensminger,^b Adelina Voukadinova,^b Alan J. Wolfe,^c  Catherine Putonti^{a,b,c,d}

^aDepartment of Biology, Loyola University Chicago, Chicago, Illinois, USA

^bBioinformatics Program, Loyola University Chicago, Chicago, Illinois, USA

^cDepartment of Microbiology and Immunology, Stritch School of Medicine, Loyola University Chicago, Maywood, Illinois, USA

^dDepartment of Computer Science, Loyola University Chicago, Chicago, Illinois, USA

ABSTRACT *Proteus mirabilis* is a Gram-negative motile and rod-shaped bacterium that is a common pathogen of the urinary tract. Here, we report the draft genome sequence of *P. mirabilis* UMB0038, which was isolated from a woman without lower urinary tract symptoms.

Proteus mirabilis is well known for its prominent flagella, allowing it to move across surfaces. Unique virulence factors such as urease production, fimbriae, iron and zinc acquisition, toxins, and biofilm formation allow it to overtake many other species with which it comes into contact, allowing it to be a common pathogen within the urinary tract (1). *P. mirabilis* is commonly seen in complicated urinary tract infection (UTI) cases that include long-term catheterization (2). Catheter-associated UTIs are frequently caused by *P. mirabilis* biofilm formation on the catheter surface, blocking urine flow. Therefore, much research has gone into methods for inhibiting biofilm formation (see reviews [3, 4]). Here, we present the draft genome sequence of a *P. mirabilis* strain, UMB0038, isolated from a catheterized urine sample obtained from a “healthy” woman without lower urinary tract symptoms.

P. mirabilis was collected as part of prior institutional review board (IRB)-approved studies (5–8) using the expanded quantitative urine culture (EQUC) protocol (8). The genus and species for this isolate were determined through matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) mass spectrometry (8) prior to storage at –80°C. From the freezer stock, the *P. mirabilis* isolate was streaked onto a Columbia nalidixic acid (CNA) plate and incubated for 24 h at 35°C in 5% CO₂. A single colony was selected from the plate, added to liquid tryptic soy broth (TSB) and incubated for 48 h at 37°C with shaking. DNA was extracted using the Qiagen DNeasy blood and tissue kit following the protocol for Gram-positive bacteria with minor modification. An adjusted amount of 230 μl of lysis buffer (180 μl of 20 mM Tris-Cl, 2 mM sodium EDTA, and 1.2% Triton X-100 and 50 μl of lysozyme) was used, with an incubation time of 10 min after the addition of buffer AL. DNA was quantified using a Qubit fluorometer and then sent to the Microbial Genome Sequencing Center (MiGS) at the University of Pittsburgh for sequencing on the Illumina NextSeq 550 platform. There the DNA was enzymatically fragmented using an Illumina tagmentation enzyme, and indices were attached using PCR. A total of 1,565,124 pairs of 150-bp reads were produced. Raw reads were trimmed using Sickle v1.33 (<https://github.com/najoshi/sickle>) and assembled using SPAdes v3.13.0 with the only-assembler option for k values of 55, 77, 99, and 127 (9). The genome coverage for the assembly was 103× and was calculated using BBMap v38.47 (<https://sourceforge.net/projects/bbmap>). Genome quality assessment and annotation were conducted with PATRIC v3.6.3 (10). The NCBI Prokaryotic Genome Annotation

Citation Gallian N, Miller-Ensminger T, Voukadinova A, Wolfe AJ, Putonti C. 2020. Draft genome sequence of *Proteus mirabilis* strain UMB0038, isolated from the female bladder. *Microbiol Resour Announc* 9:e00401-20. <https://doi.org/10.1128/MRA.00401-20>.

Editor Vincent Bruno, University of Maryland School of Medicine

Copyright © 2020 Gallian et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Catherine Putonti, cputonti@luc.edu.

Received 13 April 2020

Accepted 28 April 2020

Published 21 May 2020

Pipeline (PGAP) v4.8 (11) also was used to annotate the genome sequence. Default parameters were used for each software tool unless previously stated.

The *P. mirabilis* UMB0038 draft genome was assembled into 57 contigs. It is 3,946,388 bp long with an N_{50} value of 164,741 bp and a GC content of 38.6%. The size and GC content are similar to those of other strains of this species in GenBank. PGAP identified 3,533 protein-coding genes in the genome assembly. PHASTER (12) found three incomplete phages and one intact phage within this genome. The intact phage is of particular interest, because very few *P. mirabilis*-infecting phages are known (13) and phages are one strategy currently being explored to combat *P. mirabilis* biofilms (3). Additional analysis of this strain and genome will provide insight into how unique virulence factors contribute to UTIs.

Data availability. This whole-genome shotgun project has been deposited in GenBank under the accession no. [JAAUWO000000000](https://www.ncbi.nlm.nih.gov/nuclseq/JAAUWO000000000). The version described in this paper is the first version, JAAUWO100000000. The raw sequencing reads have been deposited in the SRA under the accession no. [SRR11441037](https://www.ncbi.nlm.nih.gov/sra/SRR11441037).

ACKNOWLEDGMENTS

This work was conducted as part of the Loyola University Chicago's Department of Biology Bacterial Genomics course. For prior patient recruitment, we acknowledge the Loyola Urinary Education and Research Collaborative and the patients who provided the samples for this study.

REFERENCES

- Schaffer J, Pearson M. 2015. *Proteus mirabilis* and urinary tract infections. *Microbiol Spectr* 3:UTI-0017–2013. <https://doi.org/10.1128/microbiolspec.UTI-0017-2013>.
- Stickler D, Ganderton L, King J, Nettleton J, Winters C. 1993. *Proteus mirabilis* biofilms and the encrustation of urethral catheters. *Urol Res* 21:407–411. <https://doi.org/10.1007/bf00300077>.
- Siddiq DM, Darouiche RO. 2012. New strategies to prevent catheter-associated urinary tract infections. *Nat Rev Urol* 9:305–314. <https://doi.org/10.1038/nrurol.2012.68>.
- Tenke P, Mezei T, Bóde I, Köves B. 2017. Catheter-associated urinary tract infections. *Eur Urol Suppl* 16:138–143. <https://doi.org/10.1016/j.eursup.2016.10.001>.
- Pearce MM, Hilt EE, Rosenfeld AB, Zilliox MJ, Thomas-White K, Fok C, Kliethermes S, Schreckenberger PC, Brubaker L, Gai X, Wolfe AJ. 2014. The female urinary microbiome: a comparison of women with and without urgency urinary incontinence. *mBio* 5:e01283-14. <https://doi.org/10.1128/mBio.01283-14>.
- Pearce MM, Zilliox MJ, Rosenfeld AB, Thomas-White KJ, Richter HE, Nager CW, Visco AG, Nygaard IE, Barber MD, Schaffer J, Moalli P, Sung VW, Smith AL, Rogers R, Nolen TL, Wallace D, Meikle SF, Gai X, Wolfe AJ, Brubaker L. 2015. The female urinary microbiome in urgency urinary incontinence. *Am J Obstet Gynecol* 213:347.e1–347.e11. <https://doi.org/10.1016/j.ajog.2015.07.009>.
- Thomas-White KJ, Hilt EE, Fok C, Pearce MM, Mueller ER, Kliethermes S, Jacobs K, Zilliox MJ, Brincat C, Price TK, Kuffel G, Schreckenberger P, Gai X, Brubaker L, Wolfe AJ. 2016. Incontinence medication response relates to the female urinary microbiota. *Int Urogynecol J* 27:723–733. <https://doi.org/10.1007/s00192-015-2847-x>.
- Hilt EE, McKinley K, Pearce MM, Rosenfeld AB, Zilliox MJ, Mueller ER, Brubaker L, Gai X, Wolfe AJ, Schreckenberger PC. 2014. Urine is not sterile: use of enhanced urine culture techniques to detect resident bacterial flora in the adult female bladder. *J Clin Microbiol* 52:871–876. <https://doi.org/10.1128/JCM.02876-13>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Wattam AR, Davis JJ, Assaf R, Boisvert S, Brettin T, Bun C, Conrad N, Dietrich EM, Disz T, Gabbard JL, Gerdes S, Henry CS, Kenyon RW, Machi D, Mao C, Nordberg EK, Olsen GJ, Murphy-Olson DE, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Vonstein V, Warren A, Xia F, Yoo H, Stevens RL. 2017. Improvements to PATRIC, the all-bacterial Bioinformatics Database and Analysis Resource Center. *Nucleic Acids Res* 45:D535–D542. <https://doi.org/10.1093/nar/gkw1017>.
- Tatusova T, DiCuccio M, Badretdin A, Chetverin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Arndt D, Grant J, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. *Nucleic Acids Res* 44:W16–W21. <https://doi.org/10.1093/nar/gkw387>.
- Alves DR, Nzakizwanayo J, Dedi C, Olympiou C, Hanin A, Kot W, Hansen L, Lametsch R, Gahan CGM, Schellenberger P, Ogilvie LA, Jones BV. 2019. Genomic and ecogenomic characterization of *Proteus mirabilis* bacteriophages. *Front Microbiol* 10:1783. <https://doi.org/10.3389/fmicb.2019.01783>.