



# Genome Sequences of Green- and Brown-Colored Strains of *Chlorobium phaeovibrioides* with Gas Vesicles

Denis S. Grouzdev,<sup>a</sup> Olga N. Lunina,<sup>a</sup> Vasil A. Gaisin,<sup>a</sup> Maria S. Krutkina,<sup>a</sup> Roman V. Baslerov,<sup>a</sup> Alexander S. Savvichev,<sup>a</sup> Vladimir M. Gorlenko<sup>a</sup>

<sup>a</sup>Research Center of Biotechnology, Russian Academy of Sciences, Moscow, Russia

**ABSTRACT** The draft genomes of green-colored *Chlorobium phaeovibrioides* GrKhr17 and brown-colored *Chlorobium phaeovibrioides* BrKhr17, green sulfur bacteria with gas vesicles isolated from Lake Bolshye Khruslomeny, are presented. These sequences contribute to genomic analyses of the *Chlorobiaceae* family that are part of ongoing research seeking to better understand their ecosystem-specific adaptations.

Meromictic lakes along the White Sea region harbor many populations of green sulfur bacteria (GSB) that offer a great opportunity for study diversity in examining the evolution of green- and brown-colored types of GSB (1–4). The green-colored strain GrKhr17 and brown-colored strain BrKhr17 of GSB (Fig. 1) were isolated from the chemocline of the meromictic Lake Bolshye Khruslomeny (Oleniy Island, Kovda Inlet, Kandalaksha Gulf, White Sea). The strains were maintained using a recently described medium (5) with sodium bicarbonate (1.5 g liter<sup>-1</sup>) and sodium sulfide (0.5 g liter<sup>-1</sup>) at 20 to 25°C in light (2,000 lx) under anaerobic conditions. The main pigment of strain GrKhr17 was bacteriochlorophyll (BChl) *d*, and the main pigment of strain BrKhr17 was BChl *e*. The pigments were determined in a 50% glycerol cell suspension using a spectrophotometer (SF-56A, OKB Spectr). A particular feature of the strains was the presence of gas vesicles, which are absent in *Chlorobium phaeovibrioides* DSM 269<sup>T</sup> and *Chlorobium phaeovibrioides* DSM 265 (6, 7).

DNA was purified from the bacterial colonies grown in the semisolid medium described earlier, and under the same conditions, using the cetyl trimethylammonium bromide (CTAB) method (8). A NEBNext Ultra DNA library prep kit (New England Biolabs, USA) was used to prepare fragment libraries for genome sequencing. Sequencing was undertaken using the Illumina HiSeq 1500 platform with single-end 250-bp reads. A total of 404,903 and 432,293 reads were obtained from GrKhr17 and BrKhr17, respectively. Low-quality reads were trimmed using a Trimmomatic v. 0.36 (9) with default settings. Subsequently, the quality-filtered reads were *de novo* assembled with SPAdes v. 3.12.0 using default settings (10). The final draft genome assembly of GrKhr17 contained 40 scaffolds, covering a total of 1,959,778 bp, with an  $L_{50}/N_{50}$  value of 4/187,559, a G+C content of 52.73%, and an average sequence coverage of 33×. The final draft genome assembly of BrKhr17 contained 67 scaffolds, covering a total of 2,094,018 bp, with an  $L_{50}/N_{50}$  value of 5/161,558, a G+C content of 53.04%, and an average sequence coverage of 31×. Identification of the protein-coding sequences and primary annotation were performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP v. 4.7) (11), which identified 1,910 genes, 1,817 protein-coding sequences, 46 pseudogenes, and 51 RNA genes for strain GrKhr17, and 1,994 genes, 1,876 protein-coding sequences, 67 pseudogenes, and 51 RNA genes for strain BrKhr17. Functional annotation of the protein-coding genes was performed using BlastKOALA

**Citation** Grouzdev DS, Lunina ON, Gaisin VA, Krutkina MS, Baslerov RV, Savvichev AS, Gorlenko VM. 2019. Genome sequences of green- and brown-colored strains of *Chlorobium phaeovibrioides* with gas vesicles. *Microbiol Resour Announc* 8:e00711-19. <https://doi.org/10.1128/MRA.00711-19>.

**Editor** David Rasko, University of Maryland School of Medicine

**Copyright** © 2019 Grouzdev et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Denis S. Grouzdev, [denisgrouzdev@gmail.com](mailto:denisgrouzdev@gmail.com).

**Received** 15 June 2019

**Accepted** 25 June 2019

**Published** 18 July 2019



**FIG 1** Color of the cultures of *Chlorobium phaeovibrioides* strains *BrKhr17* and *GrKhr17*.

(12) and supported with BLASTp (E value < 1e-20) (13) searches against the NCBI nonredundant protein database.

The 16S rRNA sequence analysis using the nucleotide BLAST (13) revealed that *GrKhr17* and *BrKhr17* share 99.93% and 99.87% similarity, respectively, with *Chlorobium phaeovibrioides* DSM 265 (GenBank accession number CP000607). The average nucleotide identity (ANI) and digital DNA-DNA hybridization (dDDH) values were calculated using the ANI calculator from the Kostas lab (<http://enve-omics.ce.gatech.edu/ani>) (14) and the Genome-to-Genome Distance Calculator (GGDC) v. 2.1 (<http://ggdc.dsmz.de/ggdc.php>) (15), respectively. In comparison with *Chlorobium phaeovibrioides* DSM 265, the ANI values of strains *GrKhr17* and *BrKhr17* were 99.1% and 99.0%, and the dDDH values were 91.7% and 90.6%, respectively. The calculated values exceeded the proposed species boundary values for species delineation (ANI < 95 to 96%, dDDH < 70%) (16), which suggests that strains *GrKhr17* and *BrKhr17* are novel strains of the known species *Chlorobium phaeovibrioides*.

The genomes of *Chlorobium phaeovibrioides* *GrKhr17* and *BrKhr17* contain the *gvp* gene cluster (17), which encodes proteins that are involved in gas vesicle biogenesis (18). The genome of *Chlorobium phaeovibrioides* *BrKhr17* contains the BChl *e* gene cluster (19), which includes BChl *e* biosynthesis genes *bciD* and *bchF3*, as well as the *cruB* gene, which is responsible for the biosynthesis of isorenieratene (20). *Chlorobium phaeovibrioides* *GrKhr17* does not possess the BChl *e* gene cluster. Both announced genomes lack genes of the *sox* system for thiosulfate oxidation. Sequencing and analysis of these bacteria revealed genomic determinants of the particular phenotype of new strains of *Chlorobium phaeovibrioides* from Arctic meromictic lakes.

**Data availability.** These whole-genome projects have been deposited in DDBJ/ENA/GenBank under the accession numbers [RXYJ0000000](https://doi.org/10.1093/nucleotide/RXYJ0000000) (*Chlorobium phaeovibrioides* *GrKhr17*) and [RXYK0000000](https://doi.org/10.1093/nucleotide/RXYK0000000) (*Chlorobium phaeovibrioides* *BrKhr17*). The versions described in this paper are RXYJ01000000 and RXYK01000000, respectively. Raw sequence reads are available under the SRA accession numbers [SRR9141222](https://doi.org/10.1093/bioinformatics/SRR9141222) (*Chlorobium phaeovibrioides* *GrKhr17*) and [SRR9141223](https://doi.org/10.1093/bioinformatics/SRR9141223) (*Chlorobium phaeovibrioides* *BrKhr17*).

## ACKNOWLEDGMENT

This study was partially supported by the Russian Foundation for Basic Research (RFBR) (research project number 17-04-01263).

## REFERENCES

- Lunina ON, Savvichev AS, Kuznetsov BB, Pimenov NV, Gorlenko VM. 2013. Anoxygenic phototrophic bacteria of the Kislo-Sladkoe stratified lake (White Sea, Kandalaksha Bay). *Microbiology* 82:815–832. <https://doi.org/10.1134/S0026261714010081>.
- Savvichev AS, Kokryatskaya NM, Zabelina SA, Rusanov II, Zakharova EE, Veslopolova EF, Lunina ON, Patutina EO, Bumazhkin BK, Gruzdev DS, Sigalevich PA, Pimenov NV, Kuznetsov BB, Gorlenko VM. 2017. Microbial processes of the carbon and sulfur cycles in an ice-covered, iron-rich meromictic lake Svetloe (Arkhangelsk region, Russia). *Environ Microbiol* 19:659–672. <https://doi.org/10.1111/1462-2920.13591>.
- Savvichev AS, Babenko VV, Lunina ON, Letarova MA, Boldyreva DI, Veslopolova EF, Demidenko NA, Kokryatskaya NM, Krasnova ED, Gaisin VA, Kostryukova ES, Gorlenko VM, Letarov AV. 2018. Sharp water column stratification with an extremely dense microbial population in a small meromictic lake, Trekhtzvetnoe. *Environ Microbiol* 20:3784–3797. <https://doi.org/10.1111/1462-2920.14384>.
- Grouzdev DS, Gaisin VA, Krutkina MS, Bryantseva IA, Lunina ON, Savvichev AS, Gorlenko VM. 2018. Genome sequence of *Prosthecochloris* sp. strain ZM and *Prosthecochloris* sp. strain ZM-2, isolated from an Arctic meromictic lake. *Microbiol Resour Announc* 7:e01415-8. <https://doi.org/10.1128/MRA.01415-18>.
- Lunina ON, Savvichev AS, Babenko VV, Boldyreva DI, Kuznetsov BB, Kolganova TV, Krasnova ED, Kokryatskaya NM, Veslopolova EF, Voronov DA, Demidenko NA, Letarova MA, Letarov AV, Gorlenko VM. 2019. Seasonal variations in the structure of an anoxygenic phototrophic bacterial community from the meromictic Lake Trekhtzvetnoe (Kandalaksha Bay, White Sea). *Microbiology* 88:100–114. <https://doi.org/10.1134/S0026261719010041>.
- Pfennig N. 1968. *Chlorobium phaeobacteroides* nov. spec. und *C. phaeovibrioides* nov. spec., zwei neue Arten der grünen Schwefelbakterien. *Arch Mikrobiol* 63:224–226. <https://doi.org/10.1007/BF00412838>.
- Imhoff JF. 2003. Phylogenetic taxonomy of the family *Chlorobiaceae* on the basis of 16S rRNA and *fmo* (Fenna-Matthews-Olson protein) gene sequences. *Int J Syst Evol Microbiol* 53:941–951. <https://doi.org/10.1099/ijs.0.02403-0>.
- Wilson K. 2001. Preparation of genomic DNA from bacteria. *Curr Protoc Mol Biol* 56:2.4.1–2.4.5. <https://doi.org/10.1002/0471142727.mb0204s56>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvermin V, Ciufu S, Li W. 2013. Prokaryotic Genome Annotation Pipeline. In *The NCBI handbook*, 2nd ed. National Center for Biotechnology Information, Bethesda, MD. <https://www.ncbi.nlm.nih.gov/books/NBK174280/>.
- Kanehisa M, Sato Y, Morishima K. 2016. BlastKOALA and GhostKOALA: KEGG tools for functional characterization of genome and metagenome sequences. *J Mol Biol* 428:726–731. <https://doi.org/10.1016/j.jmb.2015.11.006>.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
- Rodriguez-R LM, Konstantinidis KT. 2016. The enveomics collection: a toolbox for specialized analyses of microbial genomes and metagenomes. *PeerJ Prepr* 4:e1900v1. <https://doi.org/10.7287/peerj.preprints.1900v1>.
- Auch AF, von Jan M, Klenk HP, Göker M. 2010. Digital DNA-DNA hybridization for microbial species delineation by means of genome-to-genome sequence comparison. *Stand Genomic Sci* 2:117–134. <https://doi.org/10.4056/sigs.531120>.
- Chun J, Oren A, Ventosa A, Christensen H, Arahal DR, da Costa MS, Rooney AP, Yi H, Xu XW, De Meyer S, Trujillo ME. 2018. Proposed minimal standards for the use of genome data for the taxonomy of prokaryotes. *Int J Syst Evol Microbiol* 68:461–466. <https://doi.org/10.1099/ijsem.0.002516>.
- Tashiro Y, Monson RE, Ramsay JP, Salmond GP. 2016. Molecular genetic and physical analysis of gas vesicles in buoyant enterobacteria. *Environ Microbiol* 18:1264–1276. <https://doi.org/10.1111/1462-2920.13203>.
- Overmann J, Lehmann S, Pfennig N. 1991. Gas vesicle formation and buoyancy regulation in Pelodictyon phaeocathratiforme (green sulfur bacteria). *Arch Mikrobiol* 157:29–37. <https://doi.org/10.1007/BF00245331>.
- Llorens-Marès T, Liu Z, Allen LZ, Rusch DB, Craig MT, Dupont CL, Bryant DA, Casamayor EO. 2017. Speciation and ecological success in dimly lit waters: horizontal gene transfer in a green sulfur bacteria bloom unveiled by metagenomic assembly. *ISME J* 11:201–211. <https://doi.org/10.1038/ismej.2016.93>.
- Maresca JA, Romberger SP, Bryant DA. 2008. Isorenieratene biosynthesis in green sulfur bacteria requires the cooperative actions of two carotenoid cyclases. *J Bacteriol* 190:6384–6391. <https://doi.org/10.1128/JB.00758-08>.