



Complete Genome Sequence of *Serratia marcescens* Siphophage Serbin

Eric A. Williams,^a Helena Hopson,^a Andrea Rodriguez,^a Rohit Kongari,^a Rachele Bonasera,^a Adriana C. Hernandez-Morales,^a Mei Liu^a

^aCenter for Phage Technology, Texas A&M University, College Station, Texas, USA

ABSTRACT *Serratia marcescens* is an opportunistic human pathogen that is known to cause hospital-acquired respiratory and urinary tract infections. Here, we announce the complete genome sequence and the features of *S. marcescens* phage Serbin.

Serratia marcescens is a Gram-negative rod-shaped bacterium present in abundance in the environment, and infections by this bacterium are often hospital acquired and localized to the respiratory, urinary, and gastrointestinal tracts (1–3). The study of *S. marcescens* phages may help control *S. marcescens* in hospital settings.

Phage Serbin was isolated using an *S. marcescens* strain from a pond water sample collected from College Station, Texas. Nutrient broth or agar (Difco) was used to culture the host bacteria and for phage enrichment at 37°C with aeration. Phage isolation and propagation were conducted by the soft agar overlay method (4). Phage genomic DNA was prepared using a modified Promega Wizard DNA cleanup kit protocol as described previously (5). Pooled indexed DNA libraries were prepared using the Illumina TruSeq Nano low-throughput (LT) kit, and a sequence was obtained with the Illumina MiSeq platform using the MiSeq v2 500-cycle reagent kit following the manufacturer's instructions, producing 538,626 paired-end reads (250-bp read length) for the index containing the phage genome. The quality of the reads was checked in FastQC 0.11.5 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>), and reads were trimmed with FastX Toolkit 0.0.14 (http://hannonlab.cshl.edu/fastx_toolkit/download.html) and assembled in SPAdes 3.5.0 (6). The assembled genome was closed by PCR using primers (5'-CCCACCGTTAAGACTGATTAC-3' and 5'-CACCGAAGAGCACAAAGA-3') facing away from the center of the assembled contig and by Sanger sequencing of the resulting product, with the contig sequence manually corrected to match the resulting Sanger sequencing read. Protein-coding genes were predicted using GLIMMER 3.0 (7) and MetaGeneAnnotator 1.0 (8) and corrected manually if needed. The tRNA genes were predicted using ARAGORN 2.36 (9). Protein functions were predicted by comparing predicted protein sequences to the NCBI nonredundant (nr) database using BLASTp 2.2.28 (10), and conserved domains were analyzed using InterProScan 5.15-54.0 (11). All analyses were performed under default settings using the CPT Galaxy (12) and Web Apollo (13) interfaces (<https://cpt.tamu.edu>).

Serbin has a 42,882-bp genome assembled with 1,968.4-fold coverage. There were 69 protein-coding genes identified, with only 25 having predictable functions. The genome has a GC content of 51.6% and a coding density of 96.6%. Using the progressiveMAUVE algorithm (version 2.4.0) (14), Serbin shows little recognizable DNA sequence similarity to any other phage in the NCBI nucleotide database. At the protein level, phage Serbin is related to a distinct *Escherichia coli* phage group reported previously, which includes the representative *E. coli* phage 9g (GenBank accession no. [NC_024146](https://ncbi.nlm.nih.gov/nucl/NC_024146)) (15) and the more recently described four *E. coli* phages (16)

Citation Williams EA, Hopson H, Rodriguez A, Kongari R, Bonasera R, Hernandez-Morales AC, Liu M. 2019. Complete genome sequence of *Serratia marcescens* siphophage Serbin. *Microbiol Resour Announc* 8:e00422-19. <https://doi.org/10.1128/MRA.00422-19>.

Editor Simon Roux, DOE Joint Genome Institute

Copyright © 2019 Williams et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Mei Liu, meiliu@tamu.edu.

Received 8 April 2019

Accepted 18 April 2019

Published 9 May 2019

JenP2 (accession no. [KP719133](#)), JenP1 (accession no. [KP719132](#)), JenK1 (accession no. [KP719134](#)), and CAjan (accession no. [KP064094](#)). As determined by a BLASTp search (expect [E] value of $\leq 10^{-3}$), Serbin shares 22 similar proteins with these groups of *E. coli* phages, but Serbin does not have the identifiable gene cluster encoding queuosine synthesis, which is a feature shared by the phages 9g, JenP2, JenP1, JenK1, and CAjan (15, 16). Three DNA biosynthesis genes, namely thymidylate kinase, thymidylate synthase, and cytidine deaminase, were found close to one another in a set. These three genes are involved with the metabolism of nucleotides, specifically that of thymidine and cytidine (17, 18). A lysis cassette was identified, with genes coding for a holin, endolysin (*N*-acetylmuramidase), and a partially embedded i-spanin/o-spanin motif.

Data availability. The genome sequence of phage Serbin was deposited under GenBank accession no. [MK608336](#). The associated BioProject, SRA, and BioSample accession numbers are [PRJNA222858](#), [SRR8788533](#), and [SAMN11260686](#), respectively.

ACKNOWLEDGMENTS

This work was supported by funding from the National Science Foundation (awards EF-0949351 and DBI-1565146). Additional support came from the Center for Phage Technology (CPT), an Initial University Multidisciplinary Research Initiative supported by Texas A&M University and Texas AgriLife, and from the Department of Biochemistry and Biophysics at Texas A&M University.

We are grateful for the advice and support of the CPT staff.

This announcement was prepared in partial fulfillment of the requirements for BICH464 Phage Genomics, an undergraduate course at Texas A&M University.

REFERENCES

1. Hejazi A, Aucken HM, Falkiner FR. 2000. Epidemiology and susceptibility of *Serratia marcescens* in a large general hospital over an 8-year period. *J Hosp Infect* 45:42–46. <https://doi.org/10.1053/jhin.1999.0722>.
2. Khanna A, Khanna M, Aggarwal A. 2013. *Serratia marcescens*—a rare opportunistic nosocomial pathogen and measures to limit its spread in hospitalized patients. *J Clin Diagn Res* 7:243–246. <https://doi.org/10.7860/JCDR/2013/5010.2737>.
3. Jones RN. 2010. Microbial etiologies of hospital-acquired bacterial pneumonia and ventilator-associated bacterial pneumonia. *Clin Infect Dis* 51:S81–S87. <https://doi.org/10.1086/653053>.
4. Adams MK. 1959. Bacteriophages. Interscience Publishers, Inc., New York, NY.
5. Summer EJ. 2009. Preparation of a phage DNA fragment library for whole genome shotgun sequencing. *Methods Mol Biol* 502:27–46. https://doi.org/10.1007/978-1-60327-565-1_4.
6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
7. Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
8. Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <https://doi.org/10.1093/dnares/dsn027>.
9. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
10. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
11. Jones P, Binns D, Chang HY, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong SY, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>.
12. Cock PJ, Gruning BA, Paszkiwicz K, Pritchard L. 2013. Galaxy tools and workflows for sequence analysis with applications in molecular plant pathology. *PeerJ* 1:e167. <https://doi.org/10.7717/peerj.167>.
13. Lee E, Helt GA, Reese JT, Munoz-Torres MC, Childers CP, Buels RM, Stein L, Holmes IH, Elisk CG, Lewis SE. 2013. Web Apollo: a Web-based genomic annotation editing platform. *Genome Biol* 14:R93. <https://doi.org/10.1186/gb-2013-14-8-r93>.
14. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:e11147. <https://doi.org/10.1371/journal.pone.0011147>.
15. Kulikov EE, Golomidova AK, Letarova MA, Kostriukova ES, Zelenin AS, Prokhorov NS, Letarov AV. 2014. Genomic sequencing and biological characteristics of a novel *Escherichia coli* bacteriophage 9g, a putative representative of a new *Siphoviridae* genus. *Viruses* 6:5077–5092. <https://doi.org/10.3390/v6125077>.
16. Carstens AB, Kot W, Hansen LH. 2015. Complete genome sequences of four novel *Escherichia coli* bacteriophages belonging to new phage groups. *Genome Announc* 3:e00741-15. <https://doi.org/10.1128/genomeA.00741-15>.
17. Demontis S, Terao M, Brivio M, Zanotta S, Bruschi M, Garattini E. 1998. Isolation and characterization of the gene coding for human cytidine deaminase. *Biochim Biophys Acta* 1443:323–333. [https://doi.org/10.1016/S0167-4781\(98\)00235-8](https://doi.org/10.1016/S0167-4781(98)00235-8).
18. Huang SH, Tang A, Drisco B, Zhang SQ, Seeger R, Li C, Jong A. 1994. Human dTMP kinase: gene expression and enzymatic activity coinciding with cell cycle progression and cell growth. *DNA Cell Biol* 13:461–471. <https://doi.org/10.1089/dna.1994.13.461>.