



Complete Genome Sequence of *Klebsiella pneumoniae* Myophage Menlow

Heather N. Newkirk,^a Lauren Lessor,^a Jason J. Gill,^a Mei Liu^a

^aCenter for Phage Technology, Texas A&M University, College Station, Texas, USA

ABSTRACT *Klebsiella pneumoniae* is an opportunistic pathogen that has become an increasing problem in nosocomial infections. Studying phages that infect *K. pneumoniae* may lead to improvements in phage therapeutics for treating these infections. Here, the full genome sequence of Menlow, a Vi01-like phage, is introduced and described.

Klebsiella pneumoniae is a Gram-negative rod found in the normal flora of the mouth, skin, and intestines (1). Strains of *K. pneumoniae* producing the *bla*_{KPC} carbapenemase are resistant to a broad range of antibiotics and are of serious concern in hospital settings (2, 3). By identifying and characterizing phages that infect *K. pneumoniae*, new therapies and treatments may be discovered.

Phage Menlow was isolated from influent water at the Carter Creek Wastewater treatment plant in College Station, TX, against a carbapenemase-producing *K. pneumoniae* isolate of the sequence type 258 (ST258) lineage. Host bacteria were cultured on tryptic soy broth or agar (Difco) at 37°C with aeration. Phages were isolated and propagated by the soft agar overlay method (4). Phage genomic DNA was prepared using a modified Promega Wizard DNA cleanup kit protocol as described previously (5). Pooled indexed DNA libraries were prepared using the Illumina TruSeq Nano LT kit, and a sequence was obtained from the Illumina MiSeq platform using the MiSeq v2 500-cycle reagent kit following the manufacturer's instructions, producing 447,621 paired-end reads for the index containing the phage genome. Reads were quality controlled in FastQC v0.11.5 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc>), edited with FastX Toolkit v0.0.14 (http://hannonlab.cshl.edu/fastx_toolkit/index.html), and assembled in SPAdes v3.5.0 (6). The genome was closed by PCR using primers (5'-CGATGAATCCCTCTCGTTCTT-3' and 5'-CATGGGAGAGCTTCTGTACTT-3') facing off the ends of the assembled contig and Sanger sequencing of the resulting product, with the contig sequence manually corrected to match the resulting Sanger sequencing read. Protein-coding genes were predicted using GLIMMER v3.0 (7) and MetaGeneAnnotator v1.0 (8) with manual editing; tRNA genes were predicted with ARAGORN v2.36 (9). Protein functions were determined based on sequence homology detected by BLASTp v2.2.28 (10), and conserved domain searches were detected using InterProScan v5.15-5.40 (11). Nucleotide percent identity was determined by the progressiveMauve algorithm v2.4.0 (12). All analyses were conducted using default settings via Center for Phage Technology (CPT) Galaxy (13) and WebApollo (14) interfaces (cpt.tamu.edu).

The Menlow genome was assembled to 157,281 bp at 244.5-fold coverage and was predicted to encode 212 proteins and 5 tRNAs. Menlow has a coding density of 93% and a GC content of 46.4%, which is significantly lower than the GC content of *K. pneumoniae* (57%) (15). According to a BLASTp search, Menlow is largely syntenic with Vi01 (16), with 182 similar proteins at an E value of $<10^{-5}$. Menlow shows 87.3% nucleotide similarity with another *K. pneumoniae* phage, 0507-KN2-1 (GenBank accession no. [AB797215](https://genbank.ncbi.nlm.nih.gov/GenBank/AB797215)) (17). It was suggested that Menlow and other similar phages belong to a new genus called "Viunalikevirus" (18). Many genes involved in DNA

Citation Newkirk HN, Lessor L, Gill JJ, Liu M. 2019. Complete genome sequence of *Klebsiella pneumoniae* myophage Menlow. Microbiol Resour Announc 8:e00192-19. <https://doi.org/10.1128/MRA.00192-19>.

Editor John J. Dennehy, Queens College

Copyright © 2019 Newkirk et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Mei Liu, meiliu@tamu.edu.

Received 1 March 2019

Accepted 3 April 2019

Published 25 April 2019

replication and phage morphology were identified. A superinfection exclusion protein similar to that of phage P22 gp17 was identified (19). Many of the hypothetical novel genes identified are clustered around the genome's 5 tRNAs. A putative tape measure protein was identified based on the length of the protein and the presence of the alpha-helical character predicted by Jpred3 (20). The endolysin was identified as an *N*-acetylmuramidase-like lysozyme (InterPro accession no. IPR024408); however, there were no identifiable holins. A spanin complex could not be reliably identified. The TerL gene shows homology to those of phages that use headful packaging (21).

Data availability. The genome sequence of phage Menlow was deposited under GenBank accession no. [MG428990](https://ncbi.nlm.nih.gov/nucl/MG428990). The associated BioProject, SRA, and BioSample accession numbers are [PRJNA222858](https://ncbi.nlm.nih.gov/bioproject/PRJNA222858), [SRR8788475](https://ncbi.nlm.nih.gov/sra/SRR8788475), and [SAMN11259833](https://ncbi.nlm.nih.gov/biosample/SAMN11259833), respectively.

ACKNOWLEDGMENTS

This work was supported by funding from the National Science Foundation (award no. EF-0949351 and DBI-1565146) and by the National Institutes of Health (NIAID award AI121689). Additional support came from the Center for Phage Technology (CPT), an Initial University Multidisciplinary Research Initiative supported by Texas A&M University and Texas AgriLife, and from the Department of Biochemistry and Biophysics of Texas A&M University.

We thank Thomas Walsh, Weill Cornell Medical School, for the provision of bacterial isolates. We are grateful for the advice and support of the CPT staff.

This announcement was prepared in partial fulfillment of the requirements for BICH464 Phage Genomics, an undergraduate course at Texas A&M University.

REFERENCES

1. Bagley ST. 1985. Habitat association of *Klebsiella* species. *Infect Control* 6:52–58. <https://doi.org/10.1017/S0195941700062603>.
2. Sanchez GV, Master RN, Clark RB, Fyyaz M, Duvvuri P, Ekta G, Bordon J. 2013. *Klebsiella pneumoniae* antimicrobial drug resistance, United States, 1998–2010. *Emerg Infect Dis* 19:133–136. <https://doi.org/10.3201/eid1901.120310>.
3. Satlin MJ, Chen L, Patel G, Gomez-Simmonds A, Weston G, Kim AC, Seo SK, Rosenthal ME, Sperber SJ, Jenkins SG, Hamula CL, Uhlemann AC, Levi MH, Fries BC, Tang YW, Juretschko S, Rojzman AD, Hong T, Mathema B, Jacobs MR, Walsh TJ, Bonomo RA, Kreiswirth BN. 2017. Multicenter clinical and molecular epidemiological analysis of bacteremia due to carbapenem-resistant *Enterobacteriaceae* (CRE) in the CRE epicenter of the United States. *Antimicrob Agents Chemother* 61:e02349-16. <https://doi.org/10.1128/AAC.02349-16>.
4. Adams MK. 1959. Bacteriophages. Interscience Publishers, Inc., New York, NY.
5. Summer EJ. 2009. Preparation of a phage DNA fragment library for whole genome shotgun sequencing. *Methods Mol Biol* 502:27–46. https://doi.org/10.1007/978-1-60327-565-1_4.
6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
7. Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
8. Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <https://doi.org/10.1093/dnares/dsn027>.
9. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
10. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
11. Jones P, Binns D, Chang HY, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong SY, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>.
12. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:e11147. <https://doi.org/10.1371/journal.pone.0011147>.
13. Cock PJ, Gruning BA, Paszkiwicz K, Pritchard L. 2013. Galaxy tools and workflows for sequence analysis with applications in molecular plant pathology. *PeerJ* 1:e167. <https://doi.org/10.7717/peerj.167>.
14. Lee E, Helt GA, Reese JT, Munoz-Torres MC, Childers CP, Buels RM, Stein L, Holmes IH, Elisk CG, Lewis SE. 2013. Web Apollo: a Web-based genomic annotation editing platform. *Genome Biol* 14:R93. <https://doi.org/10.1186/gb-2013-14-8-r93>.
15. Hua X, Chen Q, Li X, Feng Y, Ruan Z, Yu Y. 2014. Complete genome sequence of *Klebsiella pneumoniae* sequence type 17, a multidrug-resistant strain isolated during tigecycline treatment. *Genome Announc* 2:e01337-14. <https://doi.org/10.1128/genomeA.01337-14>.
16. Pickard D, Toribio AL, Petty NK, van Tonder A, Yu L, Goulding D, Barrell B, Rance R, Harris D, Wetter M, Wain J, Choudhary J, Thomson N, Dougan G. 2010. A conserved acetyl esterase domain targets diverse bacteriophages to the Vi capsular receptor of *Salmonella enterica* serovar Typhi. *J Bacteriol* 192:5746–5754. <https://doi.org/10.1128/JB.00659-10>.
17. Hsu CR, Lin TL, Pan YJ, Hsieh PF, Wang JT. 2013. Isolation of a bacteriophage specific for a new capsular type of *Klebsiella pneumoniae* and characterization of its polysaccharide depolymerase. *PLoS One* 8:e70092. <https://doi.org/10.1371/journal.pone.0070092>.
18. Adriaenssens EM, Ackermann HW, Anany H, Blasdel B, Connerton IF, Goulding D, Griffiths MW, Hooton SP, Kutter EM, Kropinski AM, Lee JH, Maes M, Pickard D, Ryu S, Sepehrizadeh Z, Shahrabak SS, Toribio AL, Lavigne R. 2012. A suggested new bacteriophage genus: “Viunalikevirus.” *Arch Virol* 157:2035–2046. <https://doi.org/10.1007/s00705-012-1360-5>.
19. Semerjian AV, Malloy DC, Poteete AR. 1989. Genetic structure of the bacteriophage P22 PL operon. *J Mol Biol* 207:1–13. [https://doi.org/10.1016/0022-2836\(89\)90437-3](https://doi.org/10.1016/0022-2836(89)90437-3).
20. Cole C, Barber JD, Barton GJ. 2008. The Jpred 3 secondary structure prediction server. *Nucleic Acids Res* 36:W197–W201. <https://doi.org/10.1093/nar/gkn238>.
21. Casjens SR, Gilcrease EB. 2009. Determining DNA packaging strategy by analysis of the termini of the chromosomes in tailed-bacteriophage virions. *Methods Mol Biol* 502:91–111. https://doi.org/10.1007/978-1-60327-565-1_7.