



Complete Genome Sequence of a Bioactive *Pseudomonas* sp. Strain, DTU12.3, Isolated from Soil in Denmark

Pavelas Sazinas,^a May Iren Aune,^a Marie Højmark Fischer,^a Jonas Greve Lauritsen,^a Lone Gram,^a  Lars Jelsbak^a

^aDepartment of Biotechnology and Biomedicine, Technical University of Denmark, Kongens Lyngby, Denmark

ABSTRACT Here, we report the complete annotated genome sequence of a *Pseudomonas* sp. strain, DTU12.3. It was isolated from leaf-covered soil in Denmark and potentially has bioactivity against certain plant pathogens.

The *Pseudomonas* genus consists of a large number of species that are able to inhabit a diverse set of niches, from the rhizosphere to a human host. Some of the soil-dwelling pseudomonads, such as *Pseudomonas protegens* and *Pseudomonas fluorescens*, have been shown to inhibit specific bacterial and fungal phytopathogens (1, 2). In this study, we isolated and sequenced a potentially bioactive *Pseudomonas* sp. strain, DTU12.3, from leaf-covered soil in Denmark.

The DTU12.3 strain was initially isolated by diluting the collected soil sample, incubating it on selective *Pseudomonas* medium, and picking an individual fluorescent colony. 16S rRNA gene sequencing confirmed that the DTU12.3 strain belonged to the *Pseudomonas* genus. Prior to genomic DNA isolation, a liquid DTU12.3 culture was grown shaking overnight in lysogeny broth (LB) at 30°C. For Illumina sequencing, the Wizard genomic DNA purification kit (Promega) was used to isolate DNA, followed by generation of DNA libraries using a modified (half volume of each reagent) protocol of the Kapa HyperPlus library prep kit (Roche Molecular Systems) and sequencing on the Illumina MiSeq platform (300 cycles). For Nanopore sequencing, genomic DNA was isolated with the PureLink genomic DNA kit (Thermo Fisher Scientific), while DNA libraries were prepared with the rapid sequencing kit (Oxford Nanopore) and sequenced on the Nanopore MinION instrument (FLO-MIN106 flow cell). In total, 5,302,168 paired-end (2 × 150-bp) Illumina reads and 11,882 Nanopore reads (average read length, 10,881 bp; read length N_{50} , 19,417 bp) were generated. Low-quality Illumina reads were trimmed with seqtk v1.2-r94, and Nanopore reads were trimmed with Porechop v0.2.2. Unicycler v0.4.1 was used for a hybrid assembly of the DTU12.3 genome using both Illumina and Nanopore reads (3). A single circular chromosome sequence was assembled, with a size of 6,268,469 bp, G+C content of 59.46%, and average read depth of 76-fold. The genome was annotated with the NCBI Prokaryotic Genome Annotation Pipeline and was predicted to have 5,795 genes, including 5,489 protein-coding genes, 19 rRNAs, 74 tRNAs, 4 noncoding RNAs, and 209 pseudogenes (4). Based on a method by Mulet et al. (5), the BLASTN comparison of the 16S rRNA gene, *rpoD*, *rpoB*, and *gyrB* sequences across *Pseudomonas* species revealed no matches above the species identity threshold (97%). To further support this finding, the online tool JSpeciesWS was used to calculate the average nucleotide identity (ANI_b) of DTU12.3 against genomes of the 10 highest BLASTN comparison hits, and there were again no matches above the identity threshold (95%) (6). These analyses suggest that DTU12.3 could belong to a yet uncharacterized *Pseudomonas* species.

The bacterial production of secondary metabolites has been implicated in bioactivity against other bacterial and fungal species (1, 7). AntiSMASH v3.0 was used to identify eight putative secondary metabolite clusters in the genome of DTU12.3, namely, three

Citation Sazinas P, Aune MI, Fischer MH, Lauritsen JG, Gram L, Jelsbak L. 2019. Complete genome sequence of a bioactive *Pseudomonas* sp. strain, DTU12.3, isolated from soil in Denmark. Microbiol Resour Announc 8:e00121-19. <https://doi.org/10.1128/MRA.00121-19>.

Editor Vincent Bruno, University of Maryland School of Medicine

Copyright © 2019 Sazinas et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Lars Jelsbak, lj@bio.dtu.dk.

Received 1 February 2019

Accepted 16 March 2019

Published 18 April 2019

nonribosomal peptide synthetase clusters, three bacteriocins, one arylpolyene, and one unnamed cluster (8). DTU12.3 exhibits growth inhibitory activity against the bacterial phytopathogens *Xanthomonas campestris* and *Dickeya solani* *in vitro* (our unpublished data). The available genome sequence of *Pseudomonas* sp. strain DTU12.3 will potentially enable further discovery as well as functional characterization of specific genomic regions important for bioactivity against relevant plant pathogens.

Data availability. The complete genome sequence of *Pseudomonas* sp. strain DTU12.3 has been submitted to GenBank under the accession number [CP027218](https://doi.org/10.1093/nar/gkw569). Raw sequencing reads have been deposited in the Sequence Read Archive ([SRR6785587](https://doi.org/10.1093/nar/gkw569) and [SRR6785588](https://doi.org/10.1093/nar/gkw569)).

ACKNOWLEDGMENTS

This study was supported by funding from the Danish National Research Foundation (DNRF137) for the Center for Microbial Secondary Metabolites and by the Danish Council for Independent Research (6108-00300A).

REFERENCES

1. Michelsen CF, Watrous J, Glaring MA, Kersten R, Koyama N, Dorrestein PC, Stougaard P. 2015. Nonribosomal peptides, key biocontrol components for *Pseudomonas fluorescens* In5, isolated from a Greenlandic suppressive soil. *mBio* 6:e00079. <https://doi.org/10.1128/mBio.00079-15>.
2. Michavila G, Adler C, De Gregorio PR, Lami MJ, Caram Di Santo MC, Zenoff AM, de Cristobal RE, Vincent PA. 2017. *Pseudomonas protegens* CS1 from the lemon phyllosphere as a candidate for citrus canker biocontrol agent. *Plant Biol (Stuttg)* 19:608–617. <https://doi.org/10.1111/plb.12556>.
3. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
4. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44: 6614–6624. <https://doi.org/10.1093/nar/gkw569>.
5. Mulet M, Lalucat J, García-Valdés E. 2010. DNA sequence-based analysis of the *Pseudomonas* species. *Environ Microbiol* 12:1513–1530. <https://doi.org/10.1111/j.1462-2920.2010.02181.x>.
6. Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.
7. de Oliveira AG, Spago FR, Simionato AS, Navarro MO, da Silva CS, Barazetti AR, Cely MTV, Tischer CA, San Martin JAB, de Jesus Andrade CGT, Novello CR, Mello JCP, Andrade G. 2016. Bioactive organocopper compound from *Pseudomonas aeruginosa* inhibits the growth of *Xanthomonas citri* subsp. *citri*. *Front Microbiol* 7:113. <https://doi.org/10.3389/fmicb.2016.00113>.
8. Weber T, Blin K, Duddela S, Krug D, Kim HU, Brucoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. *Nucleic Acids Res* 43:W237–W243. <https://doi.org/10.1093/nar/gkv437>.