



Draft Genome Sequence of *Lecanicillium* sp. Isolate LEC01, a Fungus Capable of Hydrocarbon Degradation

Osman Radwan,^a Thusitha S. Gunasekera,^b  Oscar N. Ruiz^b

^aEnvironmental Microbiology Group, University of Dayton Research Institute, Dayton, Ohio, USA

^bFuels and Energy Branch, Aerospace Systems Directorate, Air Force Research Laboratory, Wright-Patterson AFB, Ohio, USA

ABSTRACT *Lecanicillium* sp. isolate LEC01 is adapted to grow in the presence of jet fuel, employing genes involved in the degradation of alkanes and aromatic hydrocarbons. The draft genome is estimated at 31,407,988 bp and has 9,737 proteins, 50.0% G+C content, and high similarity to *Lecanicillium* sp. strain CCF 5233.

A filamentous fungus was isolated from a hydrocarbon gas turbine fuel sample and identified as *Lecanicillium* sp. isolate LEC01, with 99% identity to *Lecanicillium* sp. strain CCF 5233 based on the 18S rRNA sequence. *Lecanicillium* species are entomopathogenic fungi secreting cell wall-degrading enzymes such as chitinases and lipases (1). Although some entomopathogenic fungi can utilize hydrocarbons (2), there has been no report of *Lecanicillium* species that are capable of metabolizing hydrocarbons. Here, we present the draft genome sequence of LEC01, highlighting genes involved in hydrocarbon adaptation and metabolism.

LEC01 was recovered from the fuel via liquid-liquid extraction (3), followed by serial dilution, plating on potato dextrose (PD) agar, and cultivation at 28°C for 3 days. The hyphal tip method (4) was used to obtain the pure isolate from discrete mycelium. Genomic DNA was isolated by the cetyltrimethylammonium bromide (CTAB) method (5) from a 1-week-old culture of LEC01 in PD broth incubated at 28°C with agitation at 200 rpm. The sequencing library was prepared using the PrepX DNA library kit and Apollo 324 next-generation sequencing (NGS) automatic library prep system (WaferGen, Fremont, CA). The ligated and indexed pre-PCR library was enriched by performing 5 cycles of PCR using the NEBNext high-fidelity 2× PCR master mix. The amplified library was purified using the Apollo PCR cleanup script and AMPure XP beads (Beckman Coulter, Brea, CA) prior to quality control (QC) analysis and DNA sequencing. The Illumina HiSeq 1000 platform was used for whole-genome shotgun (WGS) sequencing of TruSeq paired-end libraries from the LEC01 genome. WGS resulted in 54,980,257 paired-end reads with a length of 100 bp and an estimated genome coverage of 160×. Trimmomatic version 0.36 (6) was used to trim low-quality and short reads using the settings “Leading” with a threshold quality of 5, “Trailing” with a threshold quality of 5, “Slidingwindow” with an average quality of 15 across 4 bp, “Avgqual” with an average read quality of 15, and “Minlen” with a minimal length of 50 bp. The trimmed reads were *de novo* assembled using SPAdes version 3.9.1 (7) with the settings “Careful” and “Only-assembler,” resulting in 31,407,988 bp with 50.0% G+C content. The draft genome contains 794 scaffolds larger than 500 bp, with an L_{50} value of 27 contigs and an N_{50} value of 303,914 bp. The CEGMA-2.5 program (8) identified 239 out of 248 ultra-conserved eukaryotic genes, reflecting that the LEC01 draft genome is 96.37% complete. The RepeatMasker (Open-3.0) program (9) was used for masking repetitive sequences (1.28%) with a setting option of fungal species. Genes were predicted using the AUGUSTUS 3.2.1 program (10) with the parameter of the closest fungal species being *Verticillium albo-atrum*, resulting in 9,737 protein-coding genes.

Citation Radwan O, Gunasekera TS, Ruiz ON. 2019. Draft genome sequence of *Lecanicillium* sp. isolate LEC01, a fungus capable of hydrocarbon degradation. Microbiol Resour Announc 8:e01744-18. <https://doi.org/10.1128/MRA.01744-18>.

Editor Vincent Bruno, University of Maryland School of Medicine

Copyright © 2019 Radwan et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Oscar N. Ruiz, oscar.ruiz@us.af.mil.

Received 4 January 2019

Accepted 16 March 2019

Published 11 April 2019

BLASTP version 2.8.1 (11) searches against the UniProt database, *Amorphotheca resinae* reference (accession number [MADK00000000](#)), and *Aspergillus fumigatus* reference (accession number [AAHF00000000](#)) using an E value of $1e^{-5}$ identified 77.7%, 77.1%, and 77.5% of the LEC01 proteins, respectively. A local BLASTP search against the Transporter Classification Database (TCDB; <http://www.tcdb.org/>) resulted in 1,793 different transporters, including those of the major facilitator superfamily (MSF) and ATP-binding cassette (ABC), with possible involvement in the extrusion of toxic compounds. Resistant-nodulation-division (RND) efflux pump, MSF, and ABC transporters may contribute to microbial adaptation to fuel (12–15).

KEGG database and BLASTP searches identified important proteins involved in hydrocarbon degradation and insect infectivity. Chitinase, chitosanase, cutinase, hydrolyase, and lipase are examples of proteins in LEC01 that may contribute to insect host infection. The ability of LEC01 to thrive in hydrocarbon fuels is supported by the identification of proteins involved in the biodegradation of *n*-alkanes and aromatics, including cytochrome P450 alkane hydroxylase, cytochrome P450 benzoate 4-mono-oxygenase, salicylate hydroxylase, dioxygenases, succinate dehydrogenase, and catechol 1,2-dioxygenase. This genome will help in elucidating the mechanisms underlying fungal adaptation to hydrocarbons.

Data availability. This whole-genome shotgun project was deposited at DDBJ/ENA/GenBank under the accession number [NIWZ00000000](#). Raw sequences were deposited in the NCBI SRA database under accession number [SRP159109](#).

ACKNOWLEDGMENTS

This material is based on research sponsored by the AFRL/RQTF under agreement number FA8650-16-2-2605.

The U.S. Government is authorized to reproduce and distribute reprints for governmental purposes notwithstanding any copyright notation thereon. The views and conclusions contained herein are those of the authors and should not be interpreted as necessarily representing the official policies or endorsements, either expressed or implied, of the AFRL/RQTF or the U.S. Government.

REFERENCES

- Rocha-Pino Z, Marín-Cervantes M, Martínez-Archundia M, Soriano-Blancas E, Revah S, Shirai K. 2013. Morphological changes, chitinolytic enzymes and hydrophobin-like proteins as responses of *Lecanicillium lecanii* during growth with hydrocarbon. *Bioprocess Biosyst Eng* 36: 531–539. <https://doi.org/10.1007/s00449-012-0808-z>.
- Fedorak PM, Westlake D. 1986. Fungal metabolism of *n*-alkylbenzenes. *Appl Environ Microbiol* 51:435–437.
- Radwan O, Gunasekera TS, Ruiz ON. 2018. Robust multiplex quantitative polymerase chain reaction assay for universal detection of microorganisms in fuel. *Energy Fuels* 32:10530–10539. <https://doi.org/10.1021/acs.energyfuels.8b02292>.
- Hildebrand EM. 1938. Techniques for the isolation of single microorganisms. *Bot Rev* 4:628–654.
- Webb DM, Knapp SJ. 1990. DNA extraction from a previously recalcitrant plant genus. *Plant Mol Biol Rep* 8:180–185. <https://doi.org/10.1007/BF02669514>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshtkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Parra G, Bradnam K, Korf I. 2007. CEGMA: a pipeline to accurately annotate core genes in eukaryotic genomes. *Bioinformatics* 23:1061–1067. <https://doi.org/10.1093/bioinformatics/btm071>.
- Smit AFA, Hubley R, Green P. 2010. RepeatMasker Open-3.0. <http://www.repeatmasker.org>.
- Stanke M, Steinkamp R, Waack S, Morgenstern B. 2004. AUGUSTUS: a Web server for gene finding in eukaryotes. *Nucleic Acids Res* 32: W309–W312. <https://doi.org/10.1093/nar/gkh379>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
- Gunasekera TS, Bowen LL, Zhou CE, Howard-Byerly SC, Foley WS, Striebich RC, Dugan LC, Ruiz ON. 2017. Transcriptomic analyses elucidate adaptive differences of closely related strains of *Pseudomonas aeruginosa* in fuel. *Appl Environ Microbiol* 83:e03249-16. <https://doi.org/10.1128/AEM.03249-16>.
- Ruiz ON, Brown LM, Striebich RC, Smart CE, Bowen LL, Lee JS, Little BJ, Mueller SS, Gunasekera TS. 2016. Effect of conventional and alternative fuels on a marine bacterial community and the significance to bioremediation. *Energy Fuels* 30:434–444. <https://doi.org/10.1021/acs.energyfuels.5b02439>.
- Radwan O, Gunasekera TS, Ruiz ON. 2018. Draft genome sequence of *Byssochlamys* sp. isolate BYSS01, a filamentous fungus adapted to the fuel environment. *Genome Announc* 6:e00164-18. <https://doi.org/10.1128/genomeA.00164-18>.
- Radwan O, Gunasekera TS, Ruiz ON. 2018. Draft genome sequence of *Fusarium fujikuroi*, a fungus adapted to the fuel environment. *Genome Announc* 6:e01499-17. <https://doi.org/10.1128/genomeA.01499-17>.