



Complete Genome Sequence of Abscisic Acid-Metabolizing Rhizobacterium *Rhodococcus* sp. Strain P1Y

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ABSTRACT Mechanisms of microbial catabolism of phytohormone abscisic acid (ABA) are still unknown. Here, we report the complete genome sequence of ABA-utilizing *Rhodococcus* sp. strain P1Y, isolated from the rice (*Oryza sativa* L.) rhizosphere. The sequence was obtained using an approach combining Oxford Nanopore Technologies MinION and Illumina MiSeq sequence data.

Representatives of the *Rhodococcus* genus can both produce and metabolize phytohormones and their precursors (1, 2). The strain *Rhodococcus* sp. P1Y was initially isolated from the rhizosphere of rice (*Oryza sativa* L.) seedlings using a selective nutrient medium containing abscisic acid (ABA) as the sole carbon source (3). Here, it was cultivated as described previously (3). Genomic DNA was extracted and purified using the QIAamp DNA minikit (Qiagen GmbH). Paired-end and mate pair libraries were prepared using the NEBNext Ultra II DNA library kit and the Nextera mate pair library prep kit (Illumina). Sequencing was performed on an Illumina MiSeq instrument with the MiSeq reagent kit v.2 (500 cycles), which generated 2,848,635 and 100,034 reads, respectively. The paired-end reads were filtered and trimmed with PrinSeq lite v.0.20.4 (4), leaving 2,704,317 high-quality read pairs (179× genome coverage). The mate pair reads were processed with NxTrim v.0.4.2 (5), leaving 65,022 proper mate pairs (6× genome coverage).

A Nanopore library was prepared using a 1D ligation sequencing kit (SQK-LSK-108; Nanopore) and sequenced on a MinION Mk1 device. The sequencing output was 4.3 Gb (49,975 reads with a mean length of 5.5 kb and maximal length of 175.1 kb; the predicted genome coverage depth was 64×). Nanopore data assembly was performed with Canu v.1.7, which produced a single circular contig (6). Single-nucleotide polymorphisms (SNPs), short indels, and local misassemblies remaining in the assembly were fixed by Pilon v.1.22 (7) using both paired-end and mate pair Illumina data mapped onto the assembled contig by BWA-MEM (8). Multiple rounds of error correction, using the default settings of Pilon and BWA, were performed until no more errors could be fixed. The absence of large misassemblies was confirmed with mate pair data and NxRepair (9).

The complete genome sequence of *Rhodococcus* sp. strain P1Y consists of a single circular chromosome of 5,868,661 bp with a GC content of 63.19%. Annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline v.4.5 (10). A total of 5,453 genes were identified, of which 5,253 were protein-coding genes, 138 were pseudogenes, and 62 were RNA genes, of which 12 were rRNA, 47 were tRNA, and 3 were noncoding RNA (ncRNA) genes.

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The relationship of *Rhodococcus* sp. P1Y with other *Rhodococcus* spp. was assessed by calculating identities between their 16S rRNA genes. The sequences were compared to similar sequences in the NCBI database using BLAST analysis. The highest identity, at 98.62 to 98.75%, was with *Rhodococcus fascians* strain D188 (11). The average nucleotide identity between these genomes, calculated using JSpecies (12), was 74.4 to 74.7%.

Data availability. The complete genome sequence was deposited in GenBank under the accession number [CP032762](https://doi.org/10.1093/bioinformatics/btr026), corresponding to the sample accession number [SAMN10180271](https://doi.org/10.1111/j.1744-7348.2010.00439.x). The raw read files were deposited in the SRA under the accession numbers [SRX5005340](https://doi.org/10.1111/j.1744-7348.2010.00439.x), [SRX5005339](https://doi.org/10.1111/j.1744-7348.2010.00439.x), and [SRX5005338](https://doi.org/10.1111/j.1744-7348.2010.00439.x). The version described in this announcement is the first version, CP032762.1.

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REFERENCES

- Dobbelaere S, Vanderleyden J, Okon Y. 2003. Plant growth-promoting effects of diazotrophs in the rhizosphere. *Crit Rev Plant Sci* 2:107–149. <https://doi.org/10.1080/713610853>.
- Dodd IC, Zinovkina NY, Safronova VI, Belimov AA. 2010. Rhizobacterial mediation of plant hormone status. *Ann Appl Biol* 157:361–379. <https://doi.org/10.1111/j.1744-7348.2010.00439.x>.
- Belimov AA, Dodd IC, Safronova VI, Dumova VA, Shaposhnikov AI, Ladatko AG, Davies WJ. 2014. Abscisic acid metabolizing rhizobacteria decrease ABA concentrations *in planta* and alter plant growth. *Plant Physiol Biochem* 74:84–91. <https://doi.org/10.1016/j.plaphy.2013.10.032>.
- Schmieder R, Edwards R. 2011. Quality control and preprocessing of metagenomic datasets. *Bioinformatics* 27:863–864. <https://doi.org/10.1093/bioinformatics/btr026>.
- O'Connell J, Schulz-Trieglaff O, Carlson E, Hims MM, Gormley NA, Cox AJ. 2015. NxTrim: optimized trimming of Illumina mate pair reads. *Bioinformatics* 31:2035–2037. <https://doi.org/10.1093/bioinformatics/btv057>.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive *k*-mer weighting and repeat separation. *Genome Res* 27:722–736. <https://doi.org/10.1101/gr.215087.116>.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>.
- Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *arXiv* 1303:3997 [q-bio.GN]. <http://arxiv.org/abs/1303.3997>.
- Murphy RR, O'Connell J, Cox AJ, Schulz-Trieglaff O. 2015. NxRepair: error correction in *de novo* sequence assembly using Nextera mate pairs. *PeerJ* 3:e996. <https://doi.org/10.7717/peerj.996>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Stamler RA, Vereecke D, Zhang Y, Schilkey F, Devitt N, Randall JJ. 2016. Complete genome and plasmid sequences for *Rhodococcus fascians* D188 and draft sequences for *Rhodococcus* isolates PBTS 1 and PBTS 2. *Genome Announc* 4:e00495-16. <https://doi.org/10.1128/genomeA.00495-16>.
- Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci U S A* 106:19126–19131. <https://doi.org/10.1073/pnas.0906412106>.