



# Draft Genome Sequence of Multidrug-Resistant *Escherichia coli* NIVEDI-P44, Isolated from a Chicken Fecal Sample in Northeast India

Rituparna Tewari,<sup>a,b</sup> Susweta Das Mitra,<sup>a,c</sup> Sangita Das,<sup>a</sup> Sudhir Jadhao,<sup>d</sup> Gayatri Mishra,<sup>d</sup> Feroze Ganaie,<sup>a</sup> Rajeswari Shome,<sup>a</sup> Habibur Rahman,<sup>a</sup> Bibek Ranjan Shome<sup>a</sup>

<sup>a</sup>ICAR-National Institute of Veterinary Epidemiology and Disease Informatics, Bengaluru, Karnataka, India

<sup>b</sup>Department of Microbiology, Jain University, Bengaluru, Karnataka, India

<sup>c</sup>School of Basic and Applied Sciences, Dayananda Sagar University, Bengaluru, Karnataka, India

<sup>d</sup>Bionivid Pvt Ltd., Bengaluru, Karnataka, India

**ABSTRACT** We report here the draft genome sequence of a multidrug-resistant *Escherichia coli* strain (NIVEDI-P44) isolated from a chicken fecal sample. The estimated genome size is 4.76 Mb, with a G+C content of 50.65%. The genome harbors multiple antibiotic resistance genes, *bla*<sub>DHA-1</sub>, *mphA*(A), *strA*, *strB*, *dfra14*, *sul-2*, *tet*(A), and *qnrS1*.

There is worldwide concern about the emergence and rapid rise of antibiotic-resistant *Escherichia coli* strains in human and veterinary medicine in both developed and developing countries (1). Although the gut of livestock, especially poultry, is an important reservoir for drug-resistant *E. coli*, these bacterial pathogens can be transmitted to humans through direct contact, food of animal origin, and environmental routes (2, 3). Whole-genome sequencing is considered essential in the epidemiological surveillance of multidrug-resistant (MDR) strains circulating in different hosts to decipher their resistome and transmission dynamics and to gain insights into their phylogenetic and phylodynamic aspects (4). Here, we report the draft genome sequence of the MDR *E. coli* strain NIVEDI-P44, recovered from a chicken fecal sample during our molecular surveillance study of extended-spectrum- $\beta$ -lactamase- and carbapenemase-producing Gram-negative bacteria in farm animals from Northeast India. An MIC method utilizing a broth microdilution procedure revealed NIVEDI-P44 to be resistant to 7 different antibiotics (*viz.*, ampicillin, tetracycline, ciprofloxacin, cefotaxime, cefotetan, ceftazidime, and ceftriaxone). These antibiotics are of human clinical relevance and are classified as “veterinary critically important antimicrobial agents” (VCIA) in veterinary medicine.

The strain was sequenced using the Illumina HiSeq sequencing platform. The paired-end technology of Illumina platform produced a total of 12,884,022 paired-end reads of 100 bp. The next-generation sequencing quality control (NGS QC) toolkit version 2.3 (5) was used to filter high-quality data for the genome assembly. A total of 11,499,584 reads were generated and assembled using the Velvet assembler (version 1.2.10) (6), yielding 216 contigs of 4,791,462 bp and an  $N_{50}$  value of 90,342 bp. The estimated complete genome size is 4.76 Mb, with a G+C content of 50.65%. Genome annotation was performed using the Rapid Annotations using Subsystems Technology (RAST) server (7), which predicted a total of 4,611 protein-coding sequences, 73 pseudogenes, 72 tRNAs, and 3 rRNA clusters. The NIVEDI-P44 strain belongs to multi-locus sequence type (MLST) 746 (ST746). Plasmid Finder and PLACNET (8, 9) detected three plasmid sequences, *viz.*, IncN, ColpVC, and p0111 (IncHI1). Of these, p0111 harbored the *tet*(A) gene (encodes tetracycline resistance). PHAST analysis (10) detected 3 intact phages totaling 117.9 kb, which accounts for 2.46% of the total genome.

Received 23 March 2018 Accepted 24 March 2018 Published 26 April 2018

**Citation** Tewari R, Das Mitra S, Das S, Jadhao S, Mishra G, Ganaie F, Shome R, Rahman H, Shome BR. 2018. Draft genome sequence of multidrug-resistant *Escherichia coli* NIVEDI-P44, isolated from a chicken fecal sample in Northeast India. Genome Announc 6:e00205-18. <https://doi.org/10.1128/genomeA.00205-18>.

**Copyright** © 2018 Tewari et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Bibek Ranjan Shome, [brshome29@gmail.com](mailto:brshome29@gmail.com).

R.T. and S.D.M. contributed equally to this work.

Analysis by ResFinder version 2.1 (11), ARDB (12), and CARD (13) revealed the presence of the multiple antibiotic resistance genes *bla*<sub>DHA-1</sub> (class C  $\beta$ -lactamase resistance), *mph*(A) (macrolide resistance), *strA* and *strB* (streptomycin resistance), *dfpA14* (trimethoprim resistance), *sul-2* (sulfonamide resistance), and *qnrS1* (quinolone resistance).

In addition, genes encoding the major facilitator superfamily (MFS), the resistance-nodulation-division (RND) family, multidrug resistance protein A (ErmA), multidrug transporter (MdtABCD), multiple antibiotic resistance protein (Mar ABCR), and the multidrug and toxic compound extrusion (MATE) family of efflux pumps were ascertained. Further, several metal tolerance genes, namely those for nickel, copper, arsenic, cadmium, and zinc, were also identified.

The *E. coli* strains harboring such genetic resistance determinants and circulating in constantly changing environments contribute to the resistance gene pool, raising the public health threat. Therefore, diligent study of the resistome and mobilome of MDR strains widely disseminated in various environments through comparative genome evaluation will enable us to acquire information about the emergence of resistance.

**Accession number(s).** The draft genome sequences have been deposited in DDBJ/EMBL/GenBank under the accession number [LUYD00000000](https://doi.org/10.1093/nar/gkn656). The version described in this article is the first version.

## ACKNOWLEDGMENTS

This work was supported by the Department of Biotechnology, Government of India, through grants by the North Eastern Region Biotechnology Program-Animal Disease Diagnosis and Management Consortium (ADMaC) and ICAR-NICRA (National Innovation in Climate Resilient Agriculture).

We are very thankful to Arnab Sen, Principal Scientist, ICAR-Barapani, Meghalaya and his team for constant help in sample collection and processing in their laboratory.

## REFERENCES

- Cohen ML. 2000. Changing patterns of infectious disease. *Nature* 406: 762–767. <https://doi.org/10.1038/35021206>.
- Johnson JR, Russo TA. 2005. Molecular epidemiology of extraintestinal pathogenic (uropathogenic) *Escherichia coli*. *Int J Med Microbiol* 295: 383–404. <https://doi.org/10.1016/j.ijmm.2005.07.005>.
- Carattoli A. 2008. Animal reservoirs for extended spectrum  $\beta$ -lactamase producers. *Clin Microbiol Infect* 14:117–123. <https://doi.org/10.1111/j.1469-0691.2007.01851.x>.
- Azarian T, Maraqa NF, Cook RL, Johnson JA, Bailey C, Wheeler S, Nolan D, Rathore MH, Morris JG, Jr, Salemi M. 2016. Genomic epidemiology of methicillin-resistant *Staphylococcus aureus* in a neonatal intensive care unit. *PLoS One* 11:e0164397. <https://doi.org/10.1371/journal.pone.0164397>.
- Patel RK, Jain M. 2012. NGS QC toolkit: a toolkit for quality control of next generation sequencing data. *PLoS One* 7:e30619. <https://doi.org/10.1371/journal.pone.0030619>.
- Zerbino B, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <https://doi.org/10.1101/gr.074492.107>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Carattoli A, Zankari E, García-Fernández A, Voldby-Larsen M, Lund O, Villa L, Møller-Aarestrup F, Hasman H. 2014. *In silico* detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. *Antimicrob Agents Chemother* 58:3895–3903. <https://doi.org/10.1128/AAC.02412-14>.
- Lanza VF, de Toro M, Barcia PG, Mora A, Blanco J, Coque TM, Cruz F. 2014. Plasmid flux in *Escherichia coli* ST131 sublineages, analyzed by plasmid constellation network (PLACNET), a new method for plasmid reconstruction from whole genome sequences. *PLoS Genet* 10: e1004766. <https://doi.org/10.1371/journal.pgen.1004766>.
- Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. *Nucleic Acids Res* 39:W347–W352. <https://doi.org/10.1093/nar/gkr485>.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother* 67:2640–2644. <https://doi.org/10.1093/jac/dks261>.
- Liu B, Pop M. 2009. ARDB—Antibiotic Resistance Genes Database. *Nucleic Acids Res* 37:D443–D447. <https://doi.org/10.1093/nar/gkn656>.
- McArthur AG. 2013. The Comprehensive Antibiotic Resistance Database. *Antimicrob Agents Chemother* 57:3348–3357. <https://doi.org/10.1128/AAC.00419-13>.