

# Complete Genome Sequences of *Bordetella flabilis*, *Bordetella bronchialis*, and “*Bordetella pseudohinzii*”

Theodore Spilker,<sup>a</sup> Rebecca Darrah,<sup>b</sup> John J. LiPuma<sup>a</sup>

Department of Pediatrics and Communicable Diseases, University of Michigan Medical School, Ann Arbor, Michigan, USA<sup>a</sup>; Department of Genetics and Genome Sciences and Frances Payne Bolton School of Nursing, Case Western Reserve University, Cleveland, Ohio, USA<sup>b</sup>

**We report here the complete genome sequences of *Bordetella flabilis* and *Bordetella bronchialis* recovered from cultures of individuals with cystic fibrosis (CF), and “*Bordetella pseudohinzii*” recovered from a CF mouse model.**

Received 19 August 2016 Accepted 20 August 2016 Published 13 October 2016

**Citation** Spilker T, Darrah R, LiPuma JJ. 2016. Complete genome sequences of *Bordetella flabilis*, *Bordetella bronchialis*, and “*Bordetella pseudohinzii*.” *Genome Announc* 4(5): e01132-16. doi:10.1128/genomeA.01132-16.

**Copyright** © 2016 Spilker et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to John J. LiPuma, [jlipuma@umich.edu](mailto:jlipuma@umich.edu).

Individuals with cystic fibrosis (CF) are susceptible to infection of the respiratory tract with *Achromobacter* and *Bordetella* species (1–4). Correct identification of these phylogenetically closely related species may have prognostic implications and impact treatment. A recently published multilocus sequence typing scheme for *Achromobacter* revealed several novel species (5), many of which now have been taxonomically described and validly named (6–8). Sequence analysis of a fragment of *nrdA* was used to differentiate *Achromobacter* and *Bordetella* species (2–4) and identified a number of putative novel *Bordetella* species (3). Among these are the recently named *Bordetella bronchialis* and *Bordetella flabilis* (9). Another *Bordetella* genogroup, for which the name “*Bordetella pseudohinzii*” has been proposed, carries a clustered regularly interspaced short palindromic repeat (CRISPR)-associated protein 9 (Cas9) system (10). To gain further insight into the genetics of these species, we performed whole-genome sequence analysis of four strains: *B. bronchialis* AU3182, recovered from a CF patient in 2001; *B. bronchialis* AU17676, recovered from a CF patient in 2009; *B. flabilis* AU10664, recovered from a CF patient in 2006; and *B. pseudohinzii* HI4681, recovered in 2012 from bronchoalveolar lavage fluid of a C57BL/6 mouse homozygous for mutant *cfr*.

Bacteria were grown in Mueller-Hinton broth overnight at 37°C in an orbital shaker. Five milliliters of bacterial culture was pelleted and resuspended in 1 ml of 1× Tris-EDTA (TE) buffer to a concentration of ~10<sup>8</sup> CFU/ml. Genomic DNA was extracted from 350 μl of the suspension using the MagNA Pure compact nucleic acid isolation kit (Roche), according to the manufacturer’s instructions. Genomic DNA libraries were prepared using an Illumina TruSeq DNA library kit and sequenced on an Illumina HiSeq 4000 paired-end flow cell (2 × 150-bp read length, V4 chemistry). Output files containing the fastq reads were checked and edited using Trimmomatic-0.33 (11). Read correction and assembly of draft genomes were carried out using SPAdes-3.7.1 (12). Genomes were annotated using NCBI’s whole-genome shotgun (WGS) submission portal containing the automated Prokaryotic Genomic Annotation Pipeline (PGAP) option.

The contigs of each draft genome were aligned to several com-

plete *Bordetella* genomes available at NCBI, including, but not limited to, *Bordetella avium* strain 197N, *Bordetella bronchiseptica* strain 253, *Bordetella hinzii* strain H568, *Bordetella trematum* strain H044680328, and “*Bordetella* species” strain N, with Mauve version 2.4.0 (13). The reference-sorted draft genomes were manually gap filled by identifying short segments (20 to 25 bp) on the ends of two contiguous pieces that matched to both ends of a single contig of the draft genome not already included by Mauve in the alignment to the reference. These matches were verified by obtaining the longest possible perfect match on both sets of ends, checked with BLASTN for continuity, confirmed with BLASTX when possible, and checked for the appropriateness of gap distance against the reference strain. The genomes were annotated using NCBI’s whole-genome shotgun submission portal containing the automated Prokaryotic Genomic Annotation Pipeline (PGAP) option. The complete genomes, not including plasmids, ranged from 4,490,371 bp to 5,966,919 bp in length and contained 4,130 to 5,194 coding sequences (CDS) encoding proteins.

**Accession number(s).** This genome project PRJNA318508 has been deposited in GenBank under the accession numbers CP016170 to CP016173, CP016440, and CP016441.

## ACKNOWLEDGMENTS

We thank Robert Lyon, Christina McHenry, Katherine Borysko, and the University of Michigan Medical School DNA Sequencing Core Facility for their technical expertise.

This work was supported by the University of Michigan Medical School Host Microbiome Initiative and the Cystic Fibrosis Foundation.

## FUNDING INFORMATION

This work, including the efforts of Theodore Spilker and John J. LiPuma, was funded by University of Michigan (U-M). This work, including the efforts of Theodore Spilker and John J. LiPuma, was funded by Cystic Fibrosis Foundation (CF Foundation).

## REFERENCES

- Spilker T, Liwiński AA, LiPuma JJ. 2008. Identification of *Bordetella* spp. in respiratory specimens from individuals with cystic fibrosis. *Clin*

- Microbiol Infect 14:504–506. <http://dx.doi.org/10.1111/j.1469-0691.2008.01968.x>.
2. Spilker T, Vandamme P, LiPuma JJ. 2013. Identification and distribution of *Achromobacter* species in cystic fibrosis. *J Cyst Fibros* 12:298–301. <http://dx.doi.org/10.1016/j.jcf.2012.10.002>.
  3. Spilker T, Leber AL, Marcon MJ, Newton DW, Darrah R, Vandamme P, LiPuma JJ. 2014. A simplified sequence-based identification scheme for *Bordetella* reveals several putative novel species. *J Clin Microbiol* 52: 674–677. <http://dx.doi.org/10.1128/JCM.02572-13>.
  4. Coward A, Kenna DT, Perry C, Martin K, Doumith M, Turton JF. 2016. Use of *nrdA* gene sequence clustering to estimate the prevalence of different *Achromobacter* species among cystic fibrosis patients in the UK. *J Cyst Fibros* 15:479–485. <http://dx.doi.org/10.1016/j.jcf.2015.09.005>.
  5. Spilker T, Vandamme P, LiPuma JJ. 2012. A multilocus sequence typing scheme implies population structure and reveals several putative novel *Achromobacter* species. *J Clin Microbiol* 50:3010–3015. <http://dx.doi.org/10.1128/JCM.00814-12>.
  6. Vandamme P, Moore ER, Cnockaert M, De Brandt E, Svensson-Stadler L, Houf K, Spilker T, LiPuma JJ. 2013. *Achromobacter animicus* sp. nov., *Achromobacter mucicolens* sp. nov., *Achromobacter pulmonis* sp. nov. and *Achromobacter spiritinus* sp. nov., from human clinical samples. *Syst Appl Microbiol* 36:1–10. <http://dx.doi.org/10.1016/j.syapm.2012.10.003>.
  7. Vandamme P, Moore ER, Cnockaert M, Peeters C, Svensson-Stadler L, Houf K, Spilker T, LiPuma JJ. 2013. Classification of *Achromobacter* genogroups 2, 5, 7 and 14 as *Achromobacter insuavis* sp. nov., *Achromobacter aegrifaciens* sp. nov., *Achromobacter anxifer* sp. nov. and *Achromobacter dolens* sp. nov., respectively. *Syst Appl Microbiol* 36:474–482. <http://dx.doi.org/10.1016/j.syapm.2013.06.005>.
  8. Vandamme PA, Peeters C, Inganäs E, Cnockaert M, Houf K, Spilker T, Moore ER, LiPuma JJ. 2016. Taxonomic dissection of *Achromobacter denitrificans* Coenye et al. 2003 and proposal of *Achromobacter agilis* sp. nov., nom. rev., *Achromobacter pestifer* sp. nov., nom. rev., *Achromobacter kerstersii* sp. nov. and *Achromobacter deleyi* sp. nov. *Int J Syst Evol Microbiol* [Published online ahead of print June 30, 2016.]. <http://dx.doi.org/10.1099/ijsem.0.001254>.
  9. Vandamme PA, Peeters C, Cnockaert M, Inganäs E, Falsen E, Moore ER, Nunes OC, Manaia CM, Spilker T, LiPuma JJ. 2015. *Bordetella bronchialis* sp. nov., *Bordetella flabilis* sp. nov. and *Bordetella sputigena* sp. nov., isolated from human respiratory specimens, and reclassification of *Achromobacter sediminum* Zhang et al. 2014 as *Verticia sediminum* gen. nov., comb. nov. *Int J Syst Evol Microbiol* 65:3674–3682. <http://dx.doi.org/10.1099/ijsem.0.000473>.
  10. Ivanov YV, Shariat N, Register KB, Linz B, Rivera I, Hu K, Dudley EG, Harvill ET. 2015. A newly discovered *Bordetella* species carries a transcriptionally active CRISPR-Cas with a small Cas9 endonuclease. *BMC Genomics* 26:863. <http://dx.doi.org/10.1186/s12864-015-2028-9>.
  11. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <http://dx.doi.org/10.1093/bioinformatics/btu170>.
  12. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
  13. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 25: e11147. <http://dx.doi.org/10.1371/journal.pone.0011147>.