

Genome Sequence of *Aeromicrobium erythreum* NRRL B-3381, an Erythromycin-Producing Bacterium of the *Nocardioideae*

Erin A. Harrell, Eric S. Miller

Department of Plant & Microbial Biology, North Carolina State University, Raleigh, North Carolina, USA

***Aeromicrobium erythreum* NRRL B-3381 has a 3,629,239-bp circular genome that has 72% G+C content. There are at least 3,121 coding sequences (CDSs), two rRNA gene operons, and 47 tRNAs. The genome and erythromycin (*ery*) biosynthetic gene sequences provide resources for metabolic and combinatorial engineering of polyketides.**

Received 29 February 2016 Accepted 2 March 2016 Published 21 April 2016

Citation Harrell EA, Miller ES. 2016. Genome sequence of *Aeromicrobium erythreum* NRRL B-3381, an erythromycin-producing bacterium of the *Nocardioideae*. *Genome Announc* 4(2):e00300-16. doi:10.1128/genomeA.00300-16.

Copyright © 2016 Harrell and Miller. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Eric S. Miller, eric_miller@ncsu.edu.

Arthrobacter sp. strain NRRL B-3381 was part of a 1970 U.S. patent issued for an erythromycin process (1). Unlike other erythromycin processes, the NRRL B-3381 strain (isolated from Lajas Valley, Cabo Rojo, Puerto Rico) was notable for producing only erythromycin A and not the related compounds erythromycin B and C. From a large collection of industrially relevant actinobacteria, Sydney Brenner, then of the Medical Research Council (MRC) Molecular Genetics Unit, initiated a research program to genetically manipulate this nonfilamentous bacterium for polyketide combinatorial chemistry (2, 3). Since then, strain NRRL B-3381 has been taxonomically reclassified as the type genus and species *Aeromicrobium erythreum* (4), methods of plasmid transformation and gene disruption were developed (3, 5), and cosmid clones of *ery* (erythromycin) genes were isolated and sequenced (6). Although interesting metabolic manipulations of *A. erythreum* have been performed (7, 8), extensive uses of its polyketide synthase and other erythromycin biosynthesis genes have not been reported. Access to the complete genome sequence of the NRRL B-3381 strain may facilitate macrolide antibiotic development and other biotechnological uses of this and related *Actinobacteria* (9).

Total DNA was prepared using the Qiagen Gentra Puregene yeast/bact kit with overnight cultures of *A. erythreum* (collection strain designated AR18) grown at 30°C in 2xYT medium with shaking. Ten micrograms of purified DNA was processed with the Pacific Biosciences (PacBio) 10-kbp library kit in the NC State University Genomic Sciences Laboratory and then analyzed by single-molecule real-time (SMRT) RS II sequencing. Genome assembly was done with PacBio SMRT HGAP 2/Quiver using 75,640 reads with N_{50} of 5,716 bases (mean, 3,737 bases; 0.83 quality) totaling 282,723,636 bases. A total of 70,568 reads were mapped at 62.4× coverage to an assembled contig of 3,629,239 bases. No plasmid DNA was assembled. The genome is circular (by read overlap observations and PCR confirmation), has 72% G+C content, and is oriented in the GenBank file starting at 103 bases preceding the *dnaA* coding sequence (CDS).

The 3.6-Mbp genome was annotated with the NCBI Prokaryotic Genome Annotation Pipeline (10), with some manual cura-

tion. There are 3,121 CDSs, 270 pseudogenes needing further analysis, two rRNA operons (16S, 23S, and 5S), at least 47 tRNAs, and one potential transfer-messenger RNA (tmRNA). Methylation kinetics revealed m6A (N⁶-methyladenine), primarily at CTCCAG and CTGGAG (a *BpmI*-like site).

Sequences of the erythromycin-related genes, including those encoding the methyltransferase resistance enzyme (*ermR*) and the 65-kbp *ery* gene cluster, are essentially as previously reported (2, 6) (accession no. AY623658). The *ery* gene cluster includes three polyketide (6-deoxyerythronolide B) synthase modules (*eryAI—AIII*) and the methyltransferases, dehydratase, isomerase, sugar transferases, etc. leading to erythromycin A. *Aeromicrobium marinum* DSM 15272 (accession no. CM001024) encodes the most proteins currently orthologous to proteins of *A. erythreum*, and orthologs are also found from *Nocardioides simplex* VKM Ac-2033D (11).

The available *A. erythreum* NRRL B-3381 genome sequence should provide a resource for comparative and evolutionary genomics and, as suggested by Sydney Brenner 30 years ago, facilitate metabolic and combinatorial engineering of polyketide biosynthesis in a genetically tractable unicellular actinobacterium.

Nucleotide sequence accession number. The genome sequence has been deposited in GenBank with accession no. [CP011502](https://www.ncbi.nlm.nih.gov/nuccore/CP011502). The version described in this paper is the first version.

ACKNOWLEDGMENTS

We thank Cory Dashiell and Jenn Schaff of the NCSU Genomic Sciences Lab; George Yuan and Roberto Lleras of Pacific Biosciences; the NCBI Prokaryotic Genomes Annotation Pipeline group; Mark Weber of FermaLogic, Inc. for discussions; and Sydney Brenner for wit, wisdom, and innovation.

REFERENCES

- French JC, Howells JD, Anderson LE. 1970. Erythromycin process. U.S. patent 3,551,294.
- Roberts AN, Hudson GS, Brenner S. 1985. An erythromycin-resistance gene from an erythromycin-producing strain of *Arthrobacter* sp. *Gene* 35:259–270. [http://dx.doi.org/10.1016/0378-1119\(85\)90004-6](https://doi.org/10.1016/0378-1119(85)90004-6).
- Roberts AN, Barnett L, Brenner S. 1987. Transformation of *Arthrobacter*

- and studies on the transcription of the *Arthrobacter ermA* gene in *Streptomyces lividans* and *Escherichia coli*. *Biochem J* 243:431–436. <http://dx.doi.org/10.1042/bj2430431>.
4. Miller ES, Woese CR, Brenner S. 1991. Description of the erythromycin-producing bacterium *Arthrobacter* sp. strain NRRL B-3381 as *Aeromicrobium erythreum* gen. nov., sp. Nov. *Int J Syst Bacteriol* 41:363–368.
 5. Miller ES. 1991. Cloning vectors, mutagenesis, and gene disruption (*ermR*) for the erythromycin-producing bacterium *Aeromicrobium erythreum*. *Appl Environ Microbiol* 57:2758–2761.
 6. Brikun IA, Reeves AR, Cernota WH, Luu MB, Weber JM. 2004. The erythromycin biosynthetic gene cluster of *Aeromicrobium erythreum*. *J Ind Microbiol Biotechnol* 31:335–344. <http://dx.doi.org/10.1007/s10295-004-0154-5>.
 7. Reeves AR, Cernota WH, Brikun IA, Wesley RK, Weber JM. 2004. Engineering precursor flow for increased erythromycin production in *Aeromicrobium erythreum*. *Metab Eng* 6:300–312. <http://dx.doi.org/10.1016/j.ymben.2004.03.003>.
 8. Reeves AR, Seshadri R, Brikun IA, Cernota WH, Gonzalez MC, Weber JM. 2008. Knockout of the erythromycin biosynthetic cluster gene, *eryBI*, blocks isoflavone glucoside bioconversion during erythromycin fermentations in *Aeromicrobium erythreum* but not in *Saccharopolyspora erythraea*. *Appl Environ Microbiol* 74:7383–7390. <http://dx.doi.org/10.1128/AEM.01759-08>.
 9. Stackebrandt E, Rainey FA, Ward-Rainey NL. 1997. Proposal for a new hierarchic classification system, *Actinobacteria* classis nov. *Int J Syst Bacteriol* 47:479–491. <http://dx.doi.org/10.1099/00207713-47-2-479>.
 10. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufio S, Li W. 2013. Prokaryotic genome annotation pipeline. The NCBI Handbook, 2nd ed. www.ncbi.nlm.nih.gov/books/NBK174280.
 11. Shtratnikova VY, Schelkunov MI, Pekov YA, Fokina VV, Logacheva MD, Sokolov SL, Bragin EY, Ashapkin VV, Donova MV. 2015. Complete genome sequence of steroid-transforming *Nocardioides simplex* VKM Ac-2033D. *Genome Announc* 3(1):e01406-14. <http://dx.doi.org/10.1128/genomeA.01406-14>.