

Draft Genome Sequences of 14 *Escherichia coli* Phages Isolated from Cattle Slurry

R. Smith,^a M. O'Hara,^a J. L. Hobman,^b A. D. Millard^c

School of Life Sciences, University of Warwick, Coventry, United Kingdom^a; School of Biosciences, University of Nottingham, Sutton Bonington Campus, Sutton Bonington, Leicestershire, United Kingdom^b; Microbiology and Infection Unit, Warwick Medical School, University of Warwick, Coventry, United Kingdom^c

The diversity of bacteriophages in slurry from dairy cows remains largely unknown. Here, we report the draft genome sequences of 14 bacteriophages isolated from dairy cow slurry using *Escherichia coli* K-12 MG1655 as a host.

Received 2 October 2015 Accepted 4 November 2015 Published 31 December 2015

Citation Smith R, O'Hara M, Hobman JL, Millard AD. 2015. Draft genome sequences of 14 *Escherichia coli* phages isolated from cattle slurry. *Genome Announc* 3(6):e01364-15. doi:10.1128/genomeA.01364-15.

Copyright © 2015 Smith et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to A. D. Millard, a.d.millard@warwick.ac.uk.

There are approximately 1.8 million head of dairy cattle in the United Kingdom, producing a total of ~14 billion liters of milk per annum. Millions of tonnes of manure are produced by the UK dairy herd. The solid manure and liquid cow slurry produced by these animals is widely used as fertilizer and has the potential to allow the transmission of bacteria into the environment. There is concern that cattle slurry will allow the transmission of pathogenic bacteria and antibiotic resistance genes into soils (1), which ultimately may enter the human food chain. While the bacterial fraction of cattle slurry has been studied (2), the viral fraction has not. Here, we examined the diversity of bacteriophages capable of infecting *Escherichia coli*. A single slurry sample from a dairy farm slurry tank in the East Midlands of the United Kingdom was collected. The titer of phage capable of infecting *Escherichia coli* MG1655, was $4.43 \times 10^2 (\pm 1.2 \times 10^2)$ PFU/ml. A total of 30 bacteriophages were isolated, and 14 independent bacteriophage isolates were chosen at random for genome sequencing. Bacteriophage isolates were purified by a double overlay plaque assay (3). DNA was extracted from 1 mL of crude lysates using a modified phenol:chloroform extraction method (4). One nanogram of input DNA was used to prepare a genomic library for sequencing, using the Illumina Nextera XT DNA sample kit per the manufacturer's protocol (Illumina, USA). Sequencing was performed on an Illumina MiSeq instrument using the paired-end 2 × 250-bp protocol (version 2). Reads were trimmed with Sickle (5); genomes were assembled using SPAdes 3-1 with the "only-assembler" option (6). The sequence was further checked for errors by mapping reads against the resulting assembly with BWA-MEM using default settings, to correct any miscalled bases (7).

The remapping of sequences allowed the complete circularly permuted genomes to be identified (slur02, slur03, slur04, slur07, slur08, slur11, slur13, and slur14). The exact termini of the remaining phages have not been experimentally determined. Assembled genomes were annotated with Prokka version 1.11 (8) using a custom database constructed from all viral genomes available from EBI at the time. Bacteriophage genomes ranged in size from 43.9 kb (slur05) to 167.467 kb (slur08), and the G+C content ranged from 35.4% (slur14) to 54.5% (slur05). The most

common type of bacteriophage isolated belonged to the genus *T4likevirus*, with eight bacteriophage isolates (slur02, slur03, slur04, slur07, slur08, slur11, slur13, and slur14) within this genus. In comparison to the archetypal phage T4, these isolates contained far fewer endonuclease-encoding genes (e.g., *segD*, *segC*, *mobA*, and *mobB*). Homologues of the *b-gt* gene that encodes for β-glucosyltransferase, which glucosylates phage DNA and provides protection from host restriction enzymes, were also absent. Two HK578-like viruses (slur05 and slur06), along with two phages closely related to bacteriophage rv5 (slur16 and slur12), a T5-like phage (slur09) and a single phage (slur01) that had a high similarity to the previously novel enterotoxigenic *E. coli* phage Seurat (9), were also isolated. This collection of phages expands our knowledge of phages associated with cattle slurry.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession numbers LN881725 (slur01), LN881726 (slur02), LN881728 (slur03), LN881729 (slur04), LN881730 (slur05), LN881731 (slur06), LN881732 (slur07), LN881733 (slur08), LN887948 (slur09), LN881734 (slur11), LN881735 (slur12), LN881737 (slur13), LN881736 (slur14), and LN881727 (slur16).

ACKNOWLEDGMENTS

This research was supported by teaching funds from School of Life Sciences, University of Warwick (to A.D.M.), and pump priming funding from the University of Nottingham. Illumina sequencing was performed at Warwick Medical School, University of Warwick.

REFERENCES

- Sarmah AK, Meyer MT, Boxall ABA. 2006. A global perspective on the use, sales, exposure pathways, occurrence, fate and effects of veterinary antibiotics (VAs) in the environment. *Chemosphere* 65:725–759. <http://dx.doi.org/10.1016/j.chemosphere.2006.03.026>.
- Durso LM, Harhay GP, Bono JL, Smith TPL. 2011. Virulence-associated and antibiotic resistance genes of microbial populations in cattle feces analyzed using a metagenomic approach. *J Microbiol Methods* 84:278–282. <http://dx.doi.org/10.1016/j.mimet.2010.12.008>.
- Kropinski AM, Mazzocco A, Waddell TE, Lingohr E, Johnson RP. 2009. Enumeration of bacteriophages by double agar overlay plaque assay. *Methods Mol Biol* 501:69–76. http://dx.doi.org/10.1007/978-1-60327-164-6_7.

4. Clokie MRJ, Millard AD, Wilson WH, Mann NH. 2003. Encapsidation of host DNA by bacteriophages infecting marine *Synechococcus* strains. *FEMS Microbiol Ecol* 46:349–352. [http://dx.doi.org/10.1016/S0168-6496\(03\)00247-2](http://dx.doi.org/10.1016/S0168-6496(03)00247-2).
5. Joshi NA, Fass JN. 2011. Sickle: a sliding-window, adaptive, quality-based trimming tool for FastQ files. Version 1.33. <https://github.com/najoshi/sickle>.
6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
7. Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. arXiv 1303.3997. <http://arxiv.org/abs/1303.3997>.
8. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.
9. Doan DP, Lessor LE, Hernandez AC, Kutty Everett GF. 2015. Complete genome sequence of enterotoxigenic *Escherichia coli* siphophage Seurat. *Genome Announc* 3(1):e00044-15. <http://dx.doi.org/10.1128/genomeA.00044-15>.