

Draft Genome Sequences for Canadian Isolates of *Pectobacterium carotovorum* subsp. *brasiliense* with Weak Virulence on Potato

Xiang (Sean) Li,^a Kat (Xiaoli) Yuan,^a Jeff Cullis,^b C. André Lévesque,^b Wen Chen,^b Christopher T. Lewis,^b Solke H. De Boer^a

Canadian Food Inspection Agency, Charlottetown Laboratory (CFIA-CL), Charlottetown, Prince Edward Island, Canada^a; Agriculture and Agri-Food Canada (AAFC), Ottawa, Ontario, Canada^b

Pectobacterium carotovorum subsp. *brasiliense* causes soft rot and blackleg diseases on potato. Here, we report the draft genome sequences of three weakly virulent *P. carotovorum* subsp. *brasiliense* strains isolated in Canada. Analysis of these genome sequences will help to pinpoint differences in virulence among *P. carotovorum* subsp. *brasiliense* strains from tropical/subtropical and temperate regions, such as Canada and United States. A small number of key factors for adaptation to this bacterium's specific environmental niche were also evaluated.

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Address correspondence to Xiang (Sean) Li, sean.li@inspection.gc.ca.

Blackleg is an important disease of potato that causes significant losses to potato crops not only in the field but also in storage. In the past, the causal agent of the potato blackleg disease was found to be *Pectobacterium atrosepticum*. Recent studies indicated that other pectolytic bacteria also cause the blackleg disease in various potato-growing regions, for example, *Pectobacterium wasabiae* (1) and *Pectobacterium carotovorum* (2) in some temperate regions, *Dickeya solani* in European countries (3), and *P. carotovorum* subsp. *brasiliense* in Brazil (4) and some African countries (5).

P. carotovorum subsp. *brasiliense* was considered the only causal agent of potato blackleg in Brazil after an extensive survey in the Brazilian state of Rio Grande Do Sul (6), and was a major cause of potato blackleg in South Africa (5). More recently, *P. carotovorum* subsp. *brasiliense* was also found in temperate regions such as the United States (7), Canada (1), and Israel (8). In particular, De Boer et al. (1) found that Canadian isolates of *P. carotovorum* subsp. *brasiliense* were clearly less virulent than Brazilian strains in both greenhouse and field conditions. Whether or not these differences in aggressiveness could be assigned to specific genomic differences was a major trigger for this sequencing project. Comparative genomics data would provide important insight into genomic differences that differentiate the highly virulent tropical strains from temperate isolates of *P. carotovorum* subsp. *brasiliense*.

In this study, three Canadian strains (CFIA1001, 1009, and 1033) isolated from blackleg-infected potato stems were decoded using paired-end Illumina HiSeq sequencing technology with TrueSeq version 3 chemistry (National Research Council Canada, Saskatoon, Saskatchewan, Canada). In total, 2,120,303,100 bp, 1,344,825,088 bp, and 4,059,728,734 bp were obtained from 300-bp inserts to provide approximately 27×, 21×, and 37× genome coverages for strains CFIA1001, CFIA1009, and CFIA1033, respectively. After quality checking and initial *de novo* assembly using the Velvet assembler (9), the draft genome sizes for these

three strains are as follows. CFIA1001 is 4,764,478 bp comprising 28 contigs with 52.3% G+C content; CFIA1009 is 4,756,221 bp comprising 43 contigs with 51.34% G+C content; and CFIA1033 is 4,701,524 bp comprising 79 contigs with 51.4% G+C content. Annotations were conducted on the RAST server using the Glimmer 3 option (10) and it predicted 4,457, 4,442, and 4,471 protein-coding genes, including 85, 81, and 77 noncoding RNA genes for CFIA1001, CFIA1009, and CFIA1033, respectively. A number of predicted virulence-related factors, phage-related loci, motility, and chemotactic genes were identified in the genome, which may facilitate its specific pathogenicity in specific environments.

Further analysis of these strains will especially focus on environmental niche-adapted features and pathogenicity-related determinants to provide detailed insight into the genetics of ecological adaptation, virulence, and plant-pest interactions of this widely distributed pathogen.

Nucleotide sequence accession numbers. The draft genome sequences of *P. carotovorum* subsp. *brasiliense* strains CFIA1001, CFIA1009, and CFIA1033 have been deposited in GenBank under the accession numbers [JPSM00000000](https://ncbi.nlm.nih.gov/nucl/JPSM00000000), [JPSN00000000](https://ncbi.nlm.nih.gov/nucl/JPSN00000000), and [JPSO00000000](https://ncbi.nlm.nih.gov/nucl/JPSO00000000), respectively. The versions described in this paper are the first versions.

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REFERENCES

- De Boer SH, Li X, Ward LJ. 2012. *Pectobacterium* spp. associated with bacterial stem rot syndrome of potato in Canada. *Phytopathology* 102:937–947. <http://dx.doi.org/10.1094/PHYTO-04-12-0083-R>.
- De Haan EG, Dekker-Nooren TCEM, van den Bovenkamp GW,

- Speksnijder AGCL, van der Zouwen PS, van der Wolf JM. 2008. *Pectobacterium carotovorum* subsp. *carotovorum* can cause potato blackleg in temperate climates. *Eur J Plant Pathol* 122:561–569. <http://dx.doi.org/10.1007/s10658-008-9325-y>.
3. Van der Wolf JM, Nijhuis EH, Kowalewska MJ, Saddler GS, Parkinson N, Elphinstone JG, Pritchard L, Toth IK, Lojkowska E, Potrykus M, Waleron M, de Vos P, Cleenwerck I, Pirhonen M, Garlant L, Hélias V, Pothier JF, Pflüger V, Duffy B, Tsrör L, Manulis S. 2014. *Dickeya solani* sp. nov., a pectinolytic plant-pathogenic bacterium isolated from potato (*Solanum tuberosum*). *Int J Syst Evol Microbiol* 64:768–774. <http://dx.doi.org/10.1099/ijs.0.052944-0>.
 4. Duarte V, De Boer SH, Ward LJ, de Oliveira AM. 2004. Characterization of atypical *Erwinia carotovora* strains causing blackleg of potato in Brazil. *J Appl Microbiol* 96:535–545. <http://dx.doi.org/10.1111/j.1365-2672.2004.02173.x>.
 5. Van der Merwe JJ, Coutinho TA, Korsten L, van der Waals JE. 2010. *Pectobacterium carotovorum* subsp. *brasiliensis* causing blackleg on potatoes in South Africa. *Eur J Plant Pathol* 126:175–185. <http://dx.doi.org/10.1007/s10658-009-9531-2>.
 6. El Tassa SOM, Duarte V. 2004. Ocorrência de pectobactérias em tubérculos de batata-semente no estado do Rio Grande do Sul. *Fitopatol Bras* 29:620–625. <http://dx.doi.org/10.1590/S0100-41582004000600004>.
 7. Ma B, Hibbing ME, Kim HS, Reedy RM, Yedidia I, Breuer J, Breuer J, Glasner JD, Perna NT, Kelman A, Charkowski AO. 2007. Host range and molecular phylogenies of the soft rot enterobacterial genera *Pectobacterium* and *Dickeya*. *Phytopathology* 97:1150–1163. <http://dx.doi.org/10.1094/PHTO-97-9-1150>.
 8. Ali HF, Junaid M, Ahmad M, Bibi A, Ali A, Hussain S, Alam S, Shah JA. 2013. Molecular and pathogenic diversity identified among isolates of *Erwinia carotovora* subspecies *atroseptica* associated with potato blackleg and soft rot. *Pak J Bot* 45:1073–1078.
 9. Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
 10. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.