

Genomes of *Geoalkalibacter ferrihydriticus* Z-0531^T and *Geoalkalibacter subterraneus* Red1^T, Two Haloalkaliphilic Metal-Reducing *Deltaproteobacteria*

Jonathan P. Badalamenti,^a Rosa Krajmalnik-Brown,^b César I. Torres,^b Daniel R. Bond^{a,c}

BioTechnology Institute, University of Minnesota, Saint Paul, Minnesota, USA^a; Swette Center for Environmental Biotechnology, Biodesign Institute, Arizona State University, Tempe, Arizona, USA^b; Department of Microbiology, University of Minnesota, Saint Paul, Minnesota, USA^c

We sequenced and annotated genomes of two haloalkaliphilic *Deltaproteobacteria*, *Geoalkalibacter ferrihydriticus* Z-0531^T (DSM 17813) and *Geoalkalibacter subterraneus* Red1^T (DSM 23483). During assembly, we discovered that the DSMZ stock culture of *G. subterraneus* was contaminated. We reisolated *G. subterraneus* in axenic culture and redeposited it in DSMZ and JCM.

Received 14 January 2015 Accepted 2 February 2015 Published 12 March 2015

Citation Badalamenti JP, Krajmalnik-Brown R, Torres CI, Bond DR. 2015. Genomes of *Geoalkalibacter ferrihydriticus* Z-0531^T and *Geoalkalibacter subterraneus* Red1^T, two haloalkaliphilic metal-reducing *Deltaproteobacteria*. *Genome Announc* 3(2):e00039-15. doi:10.1128/genomeA.00039-15.

Copyright © 2015 Badalamenti et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Daniel R. Bond, dbond@umn.edu.

While complete genomes exist for several freshwater sediment and subsurface isolates from the *Geobacteraceae* and *Desulfuromonadaceae* families of *Deltaproteobacteria* (1), genomes from haloalkaliphilic members capable of metal reduction and electrode respiration at elevated salinity or pH (2–5) are lacking. To facilitate genetic studies of extracellular electron transfer in these haloalkaliphiles, we assembled and annotated draft and finished genomes of *Geoalkalibacter ferrihydriticus* Z-0531 and *Geoalkalibacter subterraneus* Red1, respectively.

For *G. ferrihydriticus*, we used the a5 pipeline (26 March 2013 release) (6) to assemble a 3,839,416-bp draft genome in 23 contigs at ~100× coverage (N_{50} = 602,663 bp; 57.95% GC) using 2 × 100 bp paired-end Illumina reads. More than 99% of the assembled sequence exists in 14 contigs, with the largest contig (1,394,064 bp) comprising >36% of the assembly.

Applying the same approach to *G. subterraneus*, multiple occurrences of single-copy genes and contigs with low G+C content indicated that the stock culture (5) was contaminated with a Gram-positive organism related to *Tieserella* sp. This finding was subsequently confirmed by DSMZ (S. Spring, personal communication). We reisolated *G. subterraneus* on solid medium lacking yeast extract and performed PacBio long-read sequencing according to protocols for a 20-kb insert size. SMRT bell templates were size-selected with a 4-kb cutoff using Blue Pippin electrophoresis (Sage Science). Subread filtering from 4 SMRT cells (P4-C2 chemistry, 120-min movies) yielded 1.13 Gbp of sequence (average read length = 4,760 bp; N_{50} = 6,620 bp). Assembly was performed using HGAP version 2 (7) with default parameters in SMRT Analysis version 2.1. One 400-Mbp SMRT cell was sufficient to assemble the *G. subterraneus* chromosome into one linear contig, but long reads from four SMRT cells were required to resolve three tandem copies of the rRNA operon (total length 16,597 bp) and circularize the contig. The assembly was polished to >99.999% consensus concordance (QV 50) with three successive passes through Quiver (7) at 250× coverage, and remaining indels were removed with Pilon version 1.8 (8) using 100× coverage of 2 ×

100-bp paired-end Illumina reads. The finished assembly consisted of one chromosome (3,475,523 bp) and one megaplasmid (242,122 bp) totaling 3,717,645 bp (G+C content 56.68%).

Annotation via the NCBI Prokaryotic Genome Annotation Pipeline revealed features consistent with other *Geobacteraceae* and *Desulfuromonadaceae* genomes, including a complete tricarboxylic acid cycle with a eukaryotic-like citrate synthase (9) and an abundance of putative histidine kinases (>40 in both genomes) and multiheme *c*-type cytochromes for extracellular respiration (43 and 51 in *G. ferrihydriticus* and *G. subterraneus*, respectively) (1). Notably, only *G. ferrihydriticus* contains a putative *hgcAB* gene cluster indicative of mercury methylation (GFER_06575 and GFER_06580) (10).

Nucleotide sequence accession numbers. *G. subterraneus* has been redeposited in DSMZ and JCM. Sequences have been deposited in GenBank under the accession numbers [JWJD000000000](https://ncbi.nlm.nih.gov/nucl/JWJD000000000) (*G. ferrihydriticus*) and [CP010311](https://ncbi.nlm.nih.gov/nucl/CP010311) and [CP010312](https://ncbi.nlm.nih.gov/nucl/CP010312) (*G. subterraneus* chromosome and plasmid, respectively). Raw Illumina and PacBio reads, as well as base modification data for *G. subterraneus*, have been deposited to the NCBI Sequence Read Archive under accession numbers SRX808753 and SRX808316 for *G. ferrihydriticus* and *G. subterraneus*, respectively.

ACKNOWLEDGMENTS

This work was supported by the Minnesota Environment and Natural Resources Trust Fund. Illumina sequencing was performed at the Biodesign Institute at Arizona State University.

We thank Karl Oles (Mayo Clinic Bioinformatics Core) for performing PacBio library preparation and sequencing, and Jason Smith and Scott Binford for assistance with experimental design and data analysis.

REFERENCES

- Butler JE, Young ND, Lovley DR. 2010. Evolution of electron transfer out of the cell: comparative genomics of six *Geobacter* genomes. *BMC Genomics* 11:40. <http://dx.doi.org/10.1186/1471-2164-11-40>.
- Badalamenti JP, Krajmalnik-Brown R, Torres CI. 2013. Generation of high current densities by pure cultures of anode-respiring *Geoalkalibacter*

- spp. under alkaline and saline conditions in microbial electrochemical cells. *mBio* 4:e00144-13. <http://dx.doi.org/10.1128/mBio.00144-13>.
3. Carmona-Martínez AA, Pierra M, Trably E, Bernet N. 2013. High current density via direct electron transfer by the halophilic anode respiring bacterium *Geoalkalibacter subterraneus*. *Phys Chem Chem Phys* 15: 19699–19707. <http://dx.doi.org/10.1039/c3cp54045f>.
 4. Zavarzina DG, Kolganova TV, Boulygina ES, Kostrikina NA, Tourova TP, Zavarzin GA. 2006. *Geoalkalibacter ferrihydriticus* gen. nov. sp. nov., the first alkaliphilic representative of the family *Geobacteraceae*, isolated from a soda lake. *Microbiology* 75:673–682. <http://dx.doi.org/10.1134/S0026261706060099>.
 5. Greene AC, Patel BK, Yacob S. 2009. *Geoalkalibacter subterraneus* sp. nov., an anaerobic Fe(III)- and Mn(IV)-reducing bacterium from a petroleum reservoir, and emended descriptions of the family *Desulfuromonadaceae* and the genus *Geoalkalibacter*. *Int J Syst Evol Microbiol* 59: 781–785. <http://dx.doi.org/10.1099/ijs.0.001537-0>.
 6. Tritt A, Eisen JA, Facciotti MT, Darling AE. 2012. An integrated pipeline for de novo assembly of microbial genomes. *PLoS One* 7:e42304. <http://dx.doi.org/10.1371/journal.pone.0042304>.
 7. Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <http://dx.doi.org/10.1038/nmeth.2474>.
 8. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <http://dx.doi.org/10.1371/journal.pone.0112963>.
 9. Bond DR, Mester T, Nesbø CL, Izquierdo-Lopez AV, Collart FL, Lovley DR. 2005. Characterization of citrate synthase from *Geobacter sulfurreducens* and evidence for a family of citrate synthases similar to those of eukaryotes throughout the *Geobacteraceae*. *Appl Environ Microbiol* 71: 3858–3865. <http://dx.doi.org/10.1128/AEM.71.7.3858-3865.2005>.
 10. Parks JM, Johs A, Podar M, Bridou R, Hurt RA, Smith SD, Tomanic SJ, Qian Y, Brown SD, Brandt CC, Palumbo AV, Smith JC, Wall JD, Elias DA, Liang L. 2013. The genetic basis for bacterial mercury methylation. *Science* 339:1332–1335. <http://dx.doi.org/10.1126/science.1230667>.