

# Complete Genome Sequence of Type Strain *Pasteurella multocida* subsp. *multocida* ATCC 43137

K. W. Davenport,<sup>a</sup> H. E. Daligault,<sup>a</sup> T. D. Minogue,<sup>b</sup> K. A. Bishop-Lilly,<sup>c,d</sup> D. C. Bruce,<sup>a</sup> P. S. Chain,<sup>a</sup> S. R. Coyne,<sup>b</sup> K. G. Frey,<sup>c,d</sup> J. Jaissle,<sup>b</sup> G. I. Koroleva,<sup>e</sup> J. T. Ladner,<sup>e</sup> C. C. Lo,<sup>a</sup> G. F. Palacios,<sup>e</sup> C. L. Redden,<sup>c,d</sup> M. B. Scholz,<sup>a,\*</sup> H. Teshima,<sup>a</sup> S. L. Johnson<sup>a</sup>

Los Alamos National Laboratory (LANL), Los Alamos, New Mexico, USA<sup>a</sup>; Diagnostic Systems Division (DSD), United States Army Medical Research Institute of Infectious Diseases (USAMRIID), Fort Detrick, Maryland, USA<sup>b</sup>; Naval Medical Research Center (NMRC-Frederick), Fort Detrick, Maryland, USA<sup>c</sup>; Henry M. Jackson Foundation, Bethesda, Maryland, USA<sup>d</sup>; Center for Genome Sciences (CGS), USAMRIID, Fort Detrick, Maryland, USA<sup>e</sup>

\* Present address: M. B. Scholz, Institute for Cyber-Enabled Research, Michigan State University, East Lansing, Michigan, USA.

**Soft-tissue infection by *Pasteurella multocida* in humans is usually associated with a dog- or cat-related injury, and these infections can become aggressive. We sequenced the type strain *P. multocida* subsp. *multocida* ATCC 43137 into a single closed chromosome consisting of 2,271,840 bp (40.4% G+C content), which is currently available in the NCBI GenBank under the accession number CP008918.**

Received 12 September 2014 Accepted 16 September 2014 Published 23 October 2014

**Citation** Davenport KW, Daligault HE, Minogue TD, Bishop-Lilly KA, Bruce DC, Chain PS, Coyne SR, Frey KG, Jaissle J, Koroleva GI, Ladner JT, Lo CC, Palacios GF, Redden CL, Scholz MB, Teshima H, Johnson SL. 2014. Complete genome sequence of type strain *Pasteurella multocida* subsp. *multocida* ATCC 43137. *Genome Announc.* 2(5):e01070-14. doi:10.1128/genomeA.01070-14.

**Copyright** © 2014 Davenport et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to S. L. Johnson, shannonj@lanl.gov.

*Pasteurella* is a bacterial genus composed mostly of commensals or zoonotic pathogens. However, generally as a result of scratches, bites, or licks from infected animals, several species can cause human infection. Members are small Gram-negative non-motile coccobacilli. The species comprises three subspecies (*gallinocida*, *multocida*, and *septica*), which are differentiated based on biochemical assays or PCR fingerprints (1). *P. multocida* possesses several potential virulence factors such as iron acquisition proteins, capsule lipopolysaccharide (LPS), and hemagglutinin (2, 3). *P. multocida* subsp. *multocida* ATCC 43137 is the type strain, exhibits a type-A LPS, and is commonly used as a reference strain in pathogenicity studies.

High-quality genomic DNA was extracted from a 100-ml culture of a purified isolate according to manufacturer's directions using QIAgen Genomic-tip 500. DNA was sequenced using Illumina technology. Genome assembly was performed by the Los Alamos National Laboratory (LANL) Genome Science Group, and 300-fold 100-bp paired-end (270 ± 30 bp insert) Illumina data (4) were assembled in Newbler (version 2.6), Velvet (version 1.2.08) (5), and AllPaths (version 42298) (6). Consensus sequences from all assemblers were computationally shredded and assembled with a subset of read pairs from the long-insert library using Phrap (version SPS-4.24) (7, 8). The resulting assembly was brought to closed and finished status through both manual and computational finishing efforts using Consed (9) and in-house scripts. The assembled genome sequence was corrected by mapping Illumina reads back to the final consensus sequences using Burrows-Wheeler Alignment (BWA) (10), SAMtools (11) and in-house scripts. Annotations were completed at LANL using an automated system utilizing the Ergatis workflow manager (12) and in-house scripts.

The 2.27-Mbp (40.4% G + C content) complete assembly of

*P. multocida* subsp. *multocida* ATCC 43137 assembly includes 2,076 coding sequences, 19 rRNA, and 58 tRNA sequences. Preliminary review of the annotated genome indicates resistance genes for multiple toxic metals, iron acquisition, hemagglutinin, and genes for LPS production.

**Nucleotide sequence accession number.** The complete genome assembly was deposited in GenBank under the accession number CP008918.

## ACKNOWLEDGMENTS

Funding for this effort was provided by the Defense Threat Reduction Agency's Joint Science and Technology Office (DTRA J9-CB/JSTO). This article is approved by LANL for unlimited release (LA-UR-14-25978).

The views expressed in this article are those of the authors and do not necessarily reflect the official policy or position of the Department of the Navy, Department of Defense, or the U.S. Government.

## REFERENCES

- Hunt Gerardo S, Citron DM, Claros MC, Fernandez HT, Goldstein EJC. 2001. *Pasteurella multocida* subsp. *multocida* and *P. multocida* subsp. *septica* differentiation by PCR fingerprinting and  $\alpha$ -glucosidase activity. *J. Clin. Microbiol.* 39:2558–2564. [http://dx.doi.org/10.1128/JCM.39.7.2558-2564.2001](https://doi.org/10.1128/JCM.39.7.2558-2564.2001).
- Boyce JD, Adler B. 2006. How does *Pasteurella multocida* respond to the host environment? *Curr. Opin. Microbiol.* 9:117–122. [http://dx.doi.org/10.1016/j.mib.2005.12.005](https://doi.org/10.1016/j.mib.2005.12.005).
- Harper M, Boyce JD, Adler B. 2006. *Pasteurella multocida* pathogenesis: 125 years after Pasteur. *FEMS Microbiol. Lett.* 265:1–10. [http://dx.doi.org/10.1111/j.1574-6968.2006.00442.x](https://doi.org/10.1111/j.1574-6968.2006.00442.x).
- Bennett S. 2004. Solexa Ltd. *Pharmacogenomics* 5:433–438. [http://dx.doi.org/10.1517/14622416.5.4.433](https://doi.org/10.1517/14622416.5.4.433).
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res.* 18:821–829. [http://dx.doi.org/10.1101/gr.074492.107](https://doi.org/10.1101/gr.074492.107).
- Butler J, MacCallum I, Kleber M, Shlyakhter IA, Belmonte MK, Lander

- ES, Nusbaum C, Jaffe DB. 2008. ALLPATHS: *de novo* assembly of whole-genome shotgun microreads. *Genome Res.* 18:810–820. <http://dx.doi.org/10.1101/gr.7337908>.
7. Ewing B, Green P. 1998. Base-calling of automated sequencer traces using Phred. II: error probabilities. *Genome Res.* 8:186–194.
  8. Ewing B, Hillier L, Wendl MC, Green P. 1998. Base-calling of automated sequencer traces using Phred. I: accuracy assessment. *Genome Res.* 8:175–185. <http://dx.doi.org/10.1101/gr.8.3.175>.
  9. Gordon D, Green P. 2013. *Consed*: a graphical editor for next-generation sequencing. *Bioinformatics* 29:2936–2937. <http://dx.doi.org/10.1093/bioinformatics/btt515>.
  10. Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics* 25:1754–1760. <http://dx.doi.org/10.1093/bioinformatics/btp324>.
  11. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The sequence alignment/map format and SAMtools. *Bioinformatics* 25:2078–2079. <http://dx.doi.org/10.1093/bioinformatics/btp352>.
  12. Hemmerich C, Buechlein A, Podicheti R, Revanna KV, Dong Q. 2010. An Ergatis-based prokaryotic genome annotation Web server. *Bioinformatics* 26:1122–1124. <http://dx.doi.org/10.1093/bioinformatics/btq090>.