Divergent Picobirnaviruses in Human Feces

Terry Fei Fan Ng,a,b Everardo Vega,a Nikola O. Kondov,a Christopher Markery,a Xutao Deng,a,b Nicole Gregoricusc Jan Vinjéc Eric Delwartb,a

Blood Systems Research Institute, San Francisco, California, USAa; Department of Laboratory Medicine, University of California at San Francisco, San Francisco, California, USAa; National Calicivirus Laboratory, Centers for Disease Control and Prevention, Atlanta, Georgia, USAc

The near-complete genomes of two picobirnaviruses (PBVs) in diarrheal stool samples, human picobirnaviruses D and E (HuPBV-D and -E), were genetically characterized. Their RNA-dependent RNA polymerase (RdRp) protein sequences had <66% identities to known PBVs. Due to a single nucleotide insertion, the open reading frame 2 (ORF2) in segment 1 of HuPBV-D was interrupted by a stop codon. A small stem-loop structure overlying the stop codon may result in translational readthrough into the rest of ORF2.

Picobirnaviruses (PBVs) are small nonenveloped viruses with a genome consisting of two double-stranded RNA segments first identified in rats in 1988 (1) and partly sequenced first from rabbits in 1999 and then from humans in 2000 (2, 3). The first complete genome was described in 2005 from a human fecal sample (4). PBVs have been reported in fecal and respiratory samples in humans, other mammals, reptiles, and birds (5–9). The pathogenicity of PBVs remains uncertain. Currently, fewer than 15 segment 1 sequences have been reported and only one fully characterized human PBV genome sequence has been deposited in GenBank. Most PBV sequences in GenBank include only a short conserved region of the RNA-dependent RNA polymerase (RdRp) gene in segment 2 (6).

A total of 62 fecal samples from 15 outbreaks of unexplained diarrheal disease in humans with a typical viral gastroenteritis epidemiology (10) were analyzed by viral metagenomics. The samples had previously tested negative for established enteric viral pathogens (norovirus GI, GII, and GIV, sapovirus, astrovirus, rotavirus, adenovirus, and enterovirus). Unbiased deep sequencing using an Illumina Miseq platform was performed on enriched viral particles according to previously described protocols (11, 12). BLASTx was used to identify viral sequences based on translated protein sequence similarity to virus sequences in GenBank. Two divergent PBVs were identified in samples from two individuals from separate outbreaks, and their presence was confirmed by nucleic acid reextraction, reverse transcription-PCR (RT-PCR), and Sanger sequencing. Near-complete genomes from these two human picobirnaviruses were assembled. Both viruses shared <66% identities with other PBVs in the RdRp proteins, including each other, reflecting highly distinct PBV species. To distinguish these sequences from the three existing human picobirnavirus segment 1 sequences (complete GenBank accession no. AB186897) and partial (GU968923 and AF246941), we named these human picobirnavirus D strain CDC23 (HuPBV-D-CDC23) and human picobirnavirus E strain CDC16 (HuPBV-E-CDC16).

For HuPBV-E, segment 1 was partially sequenced (2,056 bases), encoding a nearly complete capsid protein, whereas segment 2 of HuPBV-E was completely sequenced (1,717 bases). A near-complete genome was obtained for HuPBV-D, with 2,509 and 1,641 bases from segments 1 and 2, respectively, compared to 2,525 and 1,745 bases of the reference human PBV genome (NC_007026). Segment 1 normally contains 3 open reading frames (ORFs). Instead of the typical ORF2 of unknown function, the HuPBV-D ORF2 contained an extra base, resulting in an early stop codon in the middle of ORF2. We confirmed this insertion and premature ORF2 stop codon using both Illumina (30X coverage) and Sanger sequencing (4X coverage). A small stem-loop structure (5-bp stem and 9-nucleotide loop) was predicted to overlie the stop codon, indicating possible ribosomal frameshifting and translational readthrough.

With the use of RT-PCR with specific primers, only one patient from each diarrhea outbreak (among 2 and 3 tested individuals, respectively) tested positive for HuPBV-D and -E, suggesting a lack of direct association with diarrhea. A lack of association between PBV detection and diarrhea has been previously reported (3, 5), although given the wide genetic diversity of PBVs, it remains possible that certain genotypes are pathogenic in susceptible populations such as immunodeficient individuals (13).

Nucleotide sequence accession numbers. The genome sequences of HuPBV-D and -E have been submitted to GenBank under the accession numbers KJ663813 to KJ663816.

ACKNOWLEDGMENTS

We thank Sarah Ives for proofreading the manuscript.

This work was supported by NHLBI grant R01 HL105770 (to E.D.) and the Blood Systems Research Institute.

The findings and conclusions in this article are those of the authors and do not necessarily represent the views of the Centers for Disease Control and Prevention.
REFERENCES


