



# Complete Genome Sequence of *Streptomyces* Phage Sentinel

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**ABSTRACT** The *Streptomyces* genus produces over two-thirds of clinically useful, natural antibiotics. Here, we describe the isolation and genome annotation of siphophage Sentinel, which utilizes *Streptomyces* sp. strain Mg1 as a host. It has a 50,272-bp genome and 83 protein-coding genes and shows similarity to other *Streptomyces* phages in the *Arequatrovirus* genus.

*Streptomyces* spp. are Gram-positive, filamentous bacteria typically found in soil (1). They are rarely pathogenic and play an important role in the pharmaceutical industry, as members of this genus produce over two-thirds of naturally occurring antibiotics used clinically (2, 3). The uncharacterized strain *Streptomyces* sp. Mg1 has been shown to be directly antagonistic to *Bacillus subtilis* through the production of chalcomycin A, a macrolide antibiotic (4). Phage Sentinel is a novel double-stranded DNA (dsDNA) siphophage, which utilizes *Streptomyces* sp. Mg1 as a host.

Sentinel was isolated by plaque purification as described previously (5) from a topsoil sample collected in Houston, Texas, in July 2019, utilizing the host strain *Streptomyces* sp. Mg1 (6) (provided by Paul Straight, Texas A&M University). Culture conditions were 30°C and nutrient broth supplemented with 10 mM MgCl<sub>2</sub>, 8 mM Ca(NO<sub>3</sub>)<sub>2</sub>, and 0.5% glucose. DNA was purified using a Wizard DNA clean-up kit as described previously (7) and prepared as Illumina TruSeq libraries with 300-bp inserts using a Nexera Flex kit. The libraries were sequenced on an Illumina MiSeq platform with paired-end 150-bp reads using V2 (300-cycle) chemistry. The 471,318 reads were visualized with FastQC ([www.bioinformatics.babraham.ac.uk/projects/fastqc](http://www.bioinformatics.babraham.ac.uk/projects/fastqc)) and manually trimmed with FastX 0.0.14 ([http://hannonlab.cshl.edu/fastx\\_toolkit/download.html](http://hannonlab.cshl.edu/fastx_toolkit/download.html)) and then assembled using SPAdes v3.5.0 (8) into a contig with a mean coverage of 324.3×. PCR and Sanger sequencing were used to close the genome with the primers CTCCTCGGGTCATGTG (forward) and GCATCGTCATCCGAAGATC (reverse). Protein-coding genes were predicted by GLIMMER v3 (9) and MetaGeneAnnotator v1.0 (10), and detection of tRNAs was by ARAGORN v2.36 (11). Protein function was identified by conserved domain searches with InterProScan v5.33 (12), sequence similarity searches were performed with BLAST v2.9.0 (13), and transmembrane domain identification was done through TMHMM v2.0, all on default settings (14). BLAST v2.9.0 searches (accessed on 23 April 2020) were conducted against NCBI nonredundant, Swiss-Prot, and TrEMBL databases (15). ProgressiveMauve v2.4 was used to determine a genome-wide DNA sequence similarity between Sentinel and other phages (16). All annotation tools were accessed using the Galaxy platform hosted by <https://cpt.tamu.edu/galaxy-pub> (17–19). Further annotation analysis was conducted with the use of HHpred, which implements HH-suite v3 (20) software. Phage morphology was identified as that of a siphovirus (data not shown) by negatively staining samples with 2% (wt/vol) uranyl acetate and viewed by transmission electron microscopy at the Texas A&M Microscopy and Imaging Center.

The 50,272-bp genome of Sentinel has a G+C content of 66.86%, lower than the host genome G+C content of 72.17% (6). Coding density is 93.62% with a total of 83

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protein-coding genes identified. Sentinel was classified as a temperate phage due to the presence of putative integrase and immunity repressor genes (21, 22). Comparative genomics revealed a >60% nucleotide sequence similarity to 10 phages found in the *Arequatrovirus* genus. The highest nucleotide identity observed was a 65.26% similarity to *Streptomyces* phage Diane (GenBank accession number [MF766046.1](https://doi.org/10.1016/j.bjid.2012.08.014)). A putative function was assigned to 36 proteins, with the remaining 47 proteins classified as hypothetical. BLASTp analysis showed 66 putative proteins shared (E value, <0.001) between *Streptomyces* phages SqueakyClean (GenBank accession number [MF766047.1](https://doi.org/10.1016/j.bjid.2012.08.014)) and Sentinel.

**Data availability.** The genome of Sentinel was deposited in GenBank with accession number [MT701597.1](https://doi.org/10.1016/j.bjid.2012.08.014). The associated BioProject, SRA, and BioSample accession numbers are [PRJNA222858](https://doi.org/10.1016/j.bjid.2012.08.014), [SRR11574901](https://doi.org/10.1016/j.bjid.2012.08.014), and [SAMN14609633](https://doi.org/10.1016/j.bjid.2012.08.014), respectively.

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## REFERENCES

- de Lima Procópio RE, da Silva IR, Martins MK, de Azevedo JL, de Araújo JM. 2012. Antibiotics produced by *Streptomyces*. *Braz J Infect Dis* 16:466–471. <https://doi.org/10.1016/j.bjid.2012.08.014>.
- Hasani A, Kariminik A, Issazadeh K. 2014. *Streptomyces*: characteristics and their antimicrobial activities. *Int J Adv Biol Biomed Res* 2:63–75.
- Clark LC, Seipke RF, Prieto P, Willemse J, van Wezel GP, Hutchings MI, Hoskisson PA. 2013. Mammalian cell entry genes in *Streptomyces* may provide clues to the evolution of bacterial virulence. *Sci Rep* 3:1109. <https://doi.org/10.1038/srep01109>.
- Barger SR, Hoefler BC, Cubillos-Ruiz A, Russell WK, Russell DH, Straight PD. 2012. Imaging secondary metabolism of *Streptomyces* sp. Mg1 during cellular lysis and colony degradation of competing *Bacillus subtilis*. *Antonie Van Leeuwenhoek* 102:435–445. <https://doi.org/10.1007/s10482-012-9769-0>.
- van Charante F, Holtappels D, Blasdel B, Burrows B. 2019. Isolation of bacteriophages. *In* Harper D, Abedon S, Burrows B, McConville M (ed), *Bacteriophages*. Springer, Cham, Switzerland.
- Hoefler BC, Konganti K, Straight PD. 2013. De novo assembly of the *Streptomyces* sp. strain Mg1 genome using PacBio single-molecule sequencing. *Genome Announc* 1:e00535-13. <https://doi.org/10.1128/genomeA.00535-13>.
- Summer EJ. 2009. Preparation of a phage DNA fragment library for whole genome shotgun sequencing. *Methods Mol Biol* 502:27–46. [https://doi.org/10.1007/978-1-60327-565-1\\_4](https://doi.org/10.1007/978-1-60327-565-1_4).
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res* 27:4636–4641. <https://doi.org/10.1093/nar/27.23.4636>.
- Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. *DNA Res* 15:387–396. <https://doi.org/10.1093/dnares/dsn027>.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <https://doi.org/10.1093/nar/gkh152>.
- Jones P, Binns D, Chang HY, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjov M, Yong SY, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. *Bioinformatics* 30:1236–1240. <https://doi.org/10.1093/bioinformatics/btu031>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
- Krogh A, Larsson B, von Heijne G, Sonnhammer EL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 305:567–580. <https://doi.org/10.1006/jmbi.2000.4315>.
- The UniProt Consortium. 2018. UniProt: the universal protein knowledge-base. *Nucleic Acids Res* 46:2699. <https://doi.org/10.1093/nar/gky092>.
- Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:e11147. <https://doi.org/10.1371/journal.pone.0011147>.
- Ramsey J, Rasche H, Maughmer C, Criscione A, Mijalis E, Liu M, Hu JC, Young R, Gill JJ. 2020. Galaxy and Apollo as a biologist-friendly interface for high-quality cooperative phage genome annotation. *PLoS Comput Biol* 16:e1008214. <https://doi.org/10.1371/journal.pcbi.1008214>.
- Dunn NA, Unni DR, Diesh C, Munoz-Torres M, Harris NL, Yao E, Rasche H, Holmes IH, Elisk CG, Lewis SE. 2019. Apollo: democratizing genome annotation. *PLoS Comput Biol* 15:e1006790. <https://doi.org/10.1371/journal.pcbi.1006790>.
- Jalili V, Afgan E, Gu Q, Clements D, Blankenberg D, Goecks J, Taylor J, Nekrutenko A. 2020. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2020 update. *Nucleic Acids Res* 48:W395–W402. <https://doi.org/10.1093/nar/gkaa434>.
- Zimmermann L, Stephens A, Nam SZ, Rau D, Kubler J, Lozajic M, Gabler F, Soding J, Lupas AN, Alva V. 2018. A completely reimplemented MPI Bioinformatics Toolkit with a new HHpred server at its core. *J Mol Biol* 430:2237–2243. <https://doi.org/10.1016/j.jmb.2017.12.007>.
- Fogg PC, Colloms S, Rosser S, Stark M, Smith MC. 2014. New applications for phage integrases. *J Mol Biol* 426:2703–2716. <https://doi.org/10.1016/j.jmb.2014.05.014>.
- Ptashne M. 1967. Isolation of the lambda phage repressor. *Proc Natl Acad Sci U S A* 57:306–313. <https://doi.org/10.1073/pnas.57.2.306>.