

Complete Closed Genome Sequences of *Mannheimia haemolytica* Serotypes A1 and A6, Isolated from Cattle

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***Mannheimia haemolytica* is a respiratory pathogen affecting cattle and related ruminants worldwide. *M. haemolytica* is commonly associated with bovine respiratory disease complex (BRDC), a polymicrobial multifactorial disease. We present the first two complete closed genome sequences of this species, determined using an automated assembly pipeline requiring no manual finishing.**

Received 26 March 2013 Accepted 4 April 2013 Published 16 May 2013

Citation Harhay GP, Koren S, Phillippy AM, McVey DS, Kuzsak J, Clawson ML, Harhay DM, Heaton MP, Chitko-McKown CG, Smith TPL. 2013. Complete closed genome sequences of *Mannheimia haemolytica* serotypes A1 and A6, isolated from cattle. *Genome Announc*. 1(3):e00188-13. doi:10.1128/genomeA.00188-13.

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Mannheimia haemolytica is a Gram-negative rod bacterium commonly associated with severe acute hemorrhagic fibrinonecrotic bronchopneumonia (1, 2) in feedlot cattle, although it can be found in the nasopharynx of asymptomatic cattle. The bovine respiratory disease complex (BRDC) progression to bronchopneumonia results from host, pathogen, and environmental interactions that are not completely understood. Stress, viral infections, extreme weather changes, transportation, and other factors appear to predispose cattle to bronchopneumonia that is symptomatic of BRDC (3, 4). A typical BRDC outbreak occurs about 7 to 10 days after calves arrive at a feedlot. During this time, calves are at the highest risk for BRDC. Draft genome sequences of *M. haemolytica* serotypes A1 (5) and A2 (6, 7) have shown that *M. haemolytica* frequently harbors antibiotic resistance cassettes, mobile elements, and other determinants that enhance its virulence. Here, we report the first complete closed genomes of any *M. haemolytica* serotypes.

M. haemolytica strains USDA-ARS-USMARC 183 and USDA-ARS-USMARC 185 (serotypes A1 and A6, respectively) were among many *M. haemolytica* BRDC clinical case isolates that were chosen for sequencing. Genomic DNA was extracted using a Qiagen blood and cell culture DNA kit and Genomic-tip 100/G columns according to the directions of each manufacturer. Closed genomes were generated using a hybrid assembly pipeline in two steps. In step one, 50-fold coverage of Roche GS-FLX Titanium reads was used to error-correct continuous long Pacific BioScience RS sequencer reads (cLPBr) (C2 chemistry) using the pacBioToCA pipeline (8). In step two, the resulting 10,000 error-corrected cLPBr (25-fold coverage, minimum length of 6 kb) were assembled with the Celera assembler version 7 (9). Both genomes were spanned by a single contig. A dot plot of these contigs showed that the 5' end of the serotype A1 genome overlapped the 3' end by 6,021 bp with >99.8% pairwise identity, while the 5' end of the serotype A1 genome overlapped the 3' end by 3,144 bp with 99.9%

pairwise identity. For each contig of each serotype, the overlapping region was deleted from the 3' end and the two ends were joined to circularize the chromosome. GenSkew (<http://genskew.csb.univie.ac.at>) was used to localize the approximate origin of replication of the chromosome so that the base pair numbering could be reindexed to reflect the origin of replication at base pair position 1. A local instance of Do-It-Yourself Annotator (DIYA) (10) was used to annotate the circularized chromosome. The serotype A6 genome has 2,544,668 bp, a G+C content of 40.83%, 2,537 coding sequences (CDSs), 20 rRNAs, and 60 tRNAs, while the serotype A1 genome has 2,658,332 bp, a G+C content of 40.91%, 2,731 CDSs, 20 rRNAs, and 64 tRNAs.

Nucleotide sequence accession numbers. The completed genome sequences of *M. haemolytica* USDA-ARS-USMARC 183 and USDA-ARS-USMARC 185 were deposited in GenBank under accession no. CP004752 and CP004753, respectively.

ACKNOWLEDGMENTS

The contributions of G.P.H., D.S.M., M.L.C., D.M.H., M.P.H., C.G.C.-M., and T.P.L.S. were funded by the Agricultural Research Service of the United States Department of Agriculture. The contributions of S.K. and A.M.P. were funded under agreement no. HSHQDC-07-C-00020 awarded by the Department of Homeland Security (DHS) for the management and operation of the National Biodefense Analysis and Countermeasures Center (NBACC), a federally funded research and development center. The contributions of D.S.M. and J.K. were funded by the Nebraska Agriculture Experiment Station (USDA Formula Funds and UNL funds) and the Nebraska Veterinary Diagnostic Laboratory.

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REFERENCES

1. Merck & Co. Bacterial pneumonia in cattle: pneumonic pasteurellosis. The Merck veterinary manual for veterinary professionals. http://www.merckmanuals.com/vet/respiratory_system/respiratory_diseases_of_cattle/bacterial_pneumonia_in_cattle.html?qt=Pneumonic%20Pasteurellosis%26alt%3Dsh%23v3293525.
2. Rice JA, Carrasco-Medina L, Hodgins DC, Shewen PE. 2007. *Mannheimia haemolytica* and bovine respiratory disease. *Anim. Health Res. Rev.* 8:117–128.
3. Taylor JD, Fulton RW, Lehenbauer TW, Step DL, Confer AW. 2010. The epidemiology of bovine respiratory disease: what is the evidence for predisposing factors? *Can. Vet. J.* 51:1095–1102.
4. Confer AW. 2009. Update on bacterial pathogenesis in BRD. *Anim. Health Res. Rev.* 10:145–148.
5. Gioia J, Qin X, Jiang H, Clinkenbeard K, Lo R, Liu Y, Fox GE, Yerrapragada S, McLeod MP, McNeill TZ, Hemphill L, Sodergren E, Wang Q, Muzny DM, Homsy FJ, Weinstock GM, Highlander SK. 2006. The genome sequence of *Mannheimia haemolytica* A1: insights into virulence, natural competence, and *Pasteurellaceae* phylogeny. *J. Bacteriol.* 188:7257–7266.
6. Lawrence PK, Kittichotirat W, Bumgarner RE, McDermott JE, Haddon DR, Knowles DP, Srikumaran S. 2010. Genome sequences of *Mannheimia haemolytica* serotype A2: ovine and bovine isolates. *J. Bacteriol.* 192:1167–1168.
7. Lawrence PK, Kittichotirat W, McDermott JE, Bumgarner RE. 2010. A three-way comparative genomic analysis of *Mannheimia haemolytica* isolates. *BMC Genomics* 11:535.
8. Koren S, Schatz MC, Walenz BP, Martin J, Howard JT, Ganapathy G, Wang Z, Rasko DA, McCombie WR, Jarvis ED, Adam MP. 2012. Hybrid error correction and *de novo* assembly of single-molecule sequencing reads. *Nat. Biotechnol.* 30:693–700.
9. Miller JR, Delcher AL, Koren S, Venter E, Walenz BP, Brownley A, Johnson J, Li K, Mobarry C, Sutton G. 2008. Aggressive assembly of pyrosequencing reads with mates. *Bioinformatics* 24:2818–2824.
10. Stewart AC, Osborne B, Read TD. 2009. DIYA: a bacterial annotation pipeline for any genomics lab. *Bioinformatics* 25:962–963.